

PATENT
Customer No.: 32425

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:
Wendy MAURY *et al.*

Serial No.: 10/721,839

Filed: November 25, 2003

For: NOVEL ANTIVIRAL ACTIVITIES OF
PRIMATE THETA DEFENSINS AND
MAMMALIAN CATHELICIDINS

Group Art Unit: 1648

Examiner: Zachariah Lucas

Atty. Dkt. No.: IOWA:035USD1/SLH

Confirmation No.: 3197

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RESUBMITTED BRIEF ON APPEAL

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MAIL STOP APPEAL BRIEF - PATENTS

Commissioner of Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

This Resubmitted Brief on Appeal is filed in response to the Notice of Non-Compliant Appeal Brief (37 CFR 41.37) mailed on September 22, 2006, to which a response was due on October 23, 2006 since October 22, 2006 fell on a Sunday. No fees are believed due in connection with this filing. However, should any fees be due, applicants authorize the Commissioner to debit Fulbright & Jaworski L.L.P. Deposit Account No. 50-1212/IOWA:35USD1/SLH.

I. Real Party in Interest

The real party in interest of this application is the assignee, the University of Iowa Research Foundation, Iowa City, IA.

II. Related Appeals and Interferences

There are no known related appeals or interferences (see Appendix C).

III. Status of Claims

Claims 1-70 were filed with the original application, and claims 41-70 were canceled pursuant to a restriction requirement in the parent application. Claims 2-8, 16, 17 and 39 have been canceled, and claims 10-15, 25, 26 and 29-33 are withdrawn from consideration. Thus, claims 1, 9, 18-24, 27, 28, 34-38 and 40 are under examination, stand rejected, and are appealed (see Appendix A).

IV. Status of Amendments

No unentered amendments have been offered after the final rejection.

V. Summary of Invention

The claimed invention is drawn to a method for reducing the infectivity of an enveloped virus comprising contacting said virus with a first anti-viral peptide, said peptide comprising a chimeric theta defensin peptide selected from the group consisting of SEQ ID NO:31 and SEQ ID NO:32. Specification at page 3, lines 10-15 and 21-30, at page 4, lines 1-14, and at page 5, lines 1-3.

VI. Grounds on Rejection to be Reviewed on Appeal

Whether claims 1, 9, 18-24, 27, 28, 34-38 and 40 are properly rejected under 35 U.S.C. §103 as obvious over Lehrer *et al.*, WO 02/085401 (Exhibit 1).

VII. Argument

A. Standard of Review

As an initial matter, appellant notes that findings of fact and conclusions of law by the U.S. Patent and Trademark Office must be made in accordance with the Administrative Procedure Act, 5 U.S.C. § 706(A), (E), 1994, and *Dickinson v. Zurko*, 527 U.S. 150, 158 (1999). Moreover, the Federal Circuit has held that findings of fact by the Board of Patent Appeals and Interferences must be supported by “substantial evidence” within the record. *In re Gartside*, 203 F.3d 1305, 1315 (Fed. Cir. 2000). In *In re Gartside*, the Federal Circuit stated that “the ‘substantial evidence’ standard asks whether a reasonable fact finder could have arrived at the agency’s decision.” *Id.* at 1312. Accordingly, it necessarily follows that an Examiner’s position on Appeal must be supported by “substantial evidence” within the record in order to be upheld by the Board of Patent Appeals and Interferences.

B. Rejection Under 35 U.S.C. §103

Claims 1, 9, 18-24, 27, 28, 34-38 and 40 stand rejected as obvious over Lehrer *et al.* The first office action stated that a combination of Lehrer’s SEQ ID NOS: 27 and 34 would give the sequence of the present application’s SEQ ID NO:31, and that a combination of Lehrer’s SEQ ID NOS: 18 and 34 will give the sequence of the present application’s SEQ ID NO:32. The rejection is premised on the circularization of appellants’ peptides, which is contemplated by the instant specification.

The elected invention is drawn to two specific peptides, SEQ ID NOS:31 and 32, which are nowhere disclosed in the Lehrer application. The examiner recognizes this fact in that the rejection is being advanced under §103 and not §102. Thus, the reference is correctly characterized as teaching a genus of circular retrocyclin peptides comprising “two linked nonapeptides that maybe identical or different.” Lehrer at page 7, lines 8-10. Lehrer sets forth 46 different nonapeptides (SEQ ID NOS:19-64) that may be combined to form a retrocyclin. Lehrer at pages 7-17 of the Sequence Listing. Thus, if peptide “A” may be one of 46 different nonapeptides, and peptide “B” can similarly be selected from the same group, the number of members in this genus is **46 X 46**, or **2116**. The examiner has found, using appellants’ claims as a searching point, that SEQ ID NOS:31 and 32 can be identified as members of the 2000+ peptide genus described by Lehrer, and appellants do not disagree. However, this is far short of what is needed to find obviousness in this situation.

“In determining the propriety of the Patent Office case for obviousness in the first instance, it is necessary to ascertain whether or not the reference teachings would appear to be sufficient for one of ordinary skill in the relevant art having the reference before him to make the proposed substitution, combination, or other modification.” *In re Linter*, 458 F.2d 1013, 1016, 173 USPQ 560, 562 (CCPA 1972). Obviousness can only be established by combining or modifying the teachings of the prior art to produce the claimed invention where there is some teaching, suggestion, or motivation to do so found either in the references themselves or in the knowledge generally available to one of ordinary skill in the art. *In re Fine*, 837 F.2d 1071, 5 USPQ2d 1596 (Fed. Cir. 1988); *In re Jones*, 958 F.2d 347, 21 USPQ2d 1941 (Fed. Cir. 1992).

MPEP §2144.08, which is particularly instructive on this rejection, is entitled “Obviousness of Species When Prior Art Teaches Genus.” According to that section of the

MPEP, the analysis begins at the point during examination after a single prior art reference is found disclosing a genus encompassing the claimed species or subgenus. If the most relevant prior art consists of a single prior art reference disclosing a genus encompassing the claimed species, which appears to be the case here, examiner should follow the guidelines set forth therein.

Per this section, in order to determine whether the claimed species would have been obvious to one of ordinary skill in the art at the time the invention was made, the patentability of a claim to a specific compound(s) should be analyzed no differently than any other claim for purposes of §103, namely, by examining the scope and contents of the prior art, the differences between the prior art and the claims in issue, the level of skill in the pertinent art, and any evidence of secondary considerations. In particular, the fact that a claimed species is encompassed by a prior art genus is not sufficient by itself to establish a *prima facie* case of obviousness. *In re Baird*, 16 F.3d 380, 382, 29 USPQ2d 1550, 1552 (Fed. Cir. 1994) (“The fact that a claimed compound may be encompassed by a disclosed generic formula does not by itself render that compound obvious.”); *In re Jones*, 958 F.2d 347, 350, 21 USPQ2d 1941, 1943 (Fed. Cir. 1992) (Federal Circuit has “decline[d] to extract from *Merck [& Co. v. Biocraft Laboratories Inc.*, 874 F.2d 804, 10 USPQ2d 1843 (Fed. Cir. 1989)] the rule that... regardless of how broad, a disclosure of a chemical genus renders obvious any species that happens to fall within it.”). See also *In re Deuel*, 51 F.3d 1552, 1559, 34 USPQ2d 1210, 1215 (Fed. Cir. 1995).

To establish a *prima facie* case of obviousness in a genus-species chemical composition situation, it is essential that examiner find some motivation or suggestion to make the claimed invention in light of the prior art teachings. See, e.g., *In re Brouwer*, 77 F.3d 422, 425, 37 USPQ2d 1663, 1666 (Fed. Cir. 1996) (“[T]he mere possibility that one of the esters or the

active methylene group-containing compounds... could be modified or replaced such that its use would lead to the specific sulfoalkylated resin recited in claim 8 does not make the process recited in claim 8 obvious ‘unless the prior art suggested the desirability of [such a] modification’ or replacement.”) (quoting *In re Gordon*, 733 F.2d 900, 902, 221 USPQ 1125, 1127 (Fed. Cir. 1984)); *In re Vaeck*, 947 F.2d 488, 493, 20 USPQ2d 1438, 1442 (Fed. Cir. 1991) (“[A] proper analysis under §103 requires, *inter alia*, consideration of ... whether the prior art would have suggested to those of ordinary skill in the art that they should make the claimed composition or device, or carry out the claimed process.”). ***Regardless of the type of disclosure***, the prior art must provide some motivation to one of ordinary skill in the art to make the claimed invention in order to support a conclusion of obviousness. See, *e.g.*, *Vaeck*, 947 F.2d at 493, 20 USPQ2d at 1442 (A proper obviousness analysis requires consideration of “whether the prior art would also have revealed that in so making or carrying out [the claimed invention], those of ordinary skill would have a reasonable expectation of success.”); *In re Dow Chemical Co.*, 837 F.2d 469, 473, 5 USPQ2d 1529, 1531 (Fed. Cir. 1988) (“The consistent criterion for determination of obviousness is whether the prior art would have suggested to one of ordinary skill in the art that this process should be carried out and would have a reasonable likelihood of success, viewed in the light of the prior art.”); *Hodosh v. Block Drug Co.*, 786 F.2d 1136, 1143 n.5, 229 USPQ 182, 187 n.5 (Fed. Cir. 1986).

In the case of a prior art reference disclosing a genus, the examiner should make findings as to:

- (A) the structure of the disclosed prior art genus and that of any expressly described species or subgenus within the genus;
- (B) any physical or chemical properties and utilities disclosed for the genus, as well as any suggested limitations on the usefulness of the genus, and any problems alleged to be addressed by the genus;

- (C) the predictability of the technology; and
- (D) the number of species encompassed by the genus taking into consideration all of the variables possible.

As discussed above, the question boils down to whether one of ordinary skill in the relevant art would have been motivated to select the claimed species from the disclosed prior art genus. See, e.g., *Ochiai*, 71 F.3d at 1569-70, 37 USPQ2d at 1131; *Deuel*, 51 F.3d at 1557, 34 USPQ2d at 1214 (“[A] *prima facie* case of unpatentability requires that the teachings of the prior art suggest **the claimed compounds** to a person of ordinary skill in the art” (emphasis in original)); *Jones*, 958 F.2d at 351, 21 USPQ2d at 1943-44 (Fed. Cir. 1992); *Dillon*, 919 F.2d at 692, 16 USPQ2d at 1901; *In re Lahu*, 747 F.2d 703, 705, 223 USPQ 1257, 1258 (Fed. Cir. 1984) (“The prior art must provide one of ordinary skill in the art the motivation to make the proposed molecular modifications needed to arrive at the claimed compound.”). See also *In re Kemps*, 97 F.3d 1427, 1430, 40 USPQ2d 1309, 1311 (Fed. Cir. 1996) (discussing motivation to combine). The following discussion presents the PTO’s stringent requirements for such an analysis.

First, the PTO must consider the size of the prior art genus, bearing in mind that size alone cannot support an obviousness rejection. See, e.g., *Baird*, 16 F.3d at 383, 29 USPQ2d at 1552 (observing that “it is not the mere number of compounds in this limited class which is significant here but, rather, the total circumstances involved”). While there is no absolute correlation between the size of the prior art genus and a conclusion of obviousness, even a small number of genus members cannot itself create a *per se* rule of obviousness in the absence of **some** motivation to select the claimed species. See, e.g., *Deuel*, 51 F.3d at 1558-59, 34 USPQ2d at 1215 (“No particular one of these DNAs can be obvious unless there is something in the prior art to lead to the particular DNA and indicate that it should be prepared.”); *Baird*, 16 F.3d at 382-83, 29 USPQ2d at 1552; *Bell*, 991 F.2d at 784, 26 USPQ2d at 1531 (“Absent anything in the

cited prior art suggesting which of the 10^{36} possible sequences suggested by Rinderknecht corresponds to the IGF gene, the PTO has not met its burden of establishing that the prior art would have suggested the claimed sequences.”); *In re Ruschig*, 343 F.2d 965, 974, 145 USPQ 274, 282 (CCPA 1965) (Rejection of claimed compound in light of prior art genus based on *Petering* is not appropriate where the prior art does not disclose a small recognizable class of compounds with common properties.). Here, there is a large genus, and more importantly, no discussion of why the claimed species would have been selected.

Other relevant factors noticeably overlooked in the action are the number of variables which must be selected or modified, and the nature and significance of the differences between the prior art and the claimed invention. See, e.g., *In re Jones*, 958 F.2d 347, 350, 21 USPQ2d 1941, 1943 (Fed. Cir. 1992) (reversing obviousness rejection of novel dicamba salt with acyclic structure over broad prior art genus encompassing claimed salt, where disclosed examples of genus were dissimilar in structure, lacking an ether linkage or being cyclic); *In re Susi*, 440 F.2d 442, 445, 169 USPQ 423, 425 (CCPA 1971) (the difference from the particularly preferred subgenus of the prior art was a hydroxyl group, a difference conceded by applicant “to be of little importance”). In the area of biotechnology, of which the present invention is an example, an exemplified species may differ from a claimed species by a conservative or non-conservative substitution, although at some locations even a conservative substitution may not be permitted. For example, the gain or loss of even one methyl group can destabilize the structure if close packing is required in the interior of domains. James Darnell *et al.*, *Molecular Cell Biology* (3rd ed. 1998; Exhibit 2). Thus, it is incumbent upon the examiner here to show *some* evidence that would motivate one of skill the art to *select* SEQ ID NO:31 or 32 from the 2116 members of this genus.

The final office action feebly counters this argument by simply stating that each and every one of the various peptides encompassed by the reference's generic disclosure would be obvious. This statement is flawed on its face, as it treats the reference as having disclosed all 2116 peptides when it did *not*. Moreover, it constitutes an impermissible leap to a conclusion of obviousness that avoids the very clear mandate set forth by the MPEP and the controlling case law, excerpted above, to point to guidance in the cited art to arrive at the claimed invention. Again, at the risk of belaboring the issue, Lehrer's disclosure is *generic*, and there is no precise disclosure of either SEQ ID NO:31 or 32 in that reference – indeed, were there such a disclosure, appellants would be facing a §102 rejection. But they are *not* disclosed, and the rejection is not advanced under §102, so the examiner *must* provide a proper analysis under §103, including motivation to select the undisclosed species, which has not been done.

Also relevant, and notably unaddressed in the action, is the general predictability of the technology. See, *e.g.*, *Dillon*, 919 F.2d at 692-97, 16 USPQ2d at 1901-05; *In re Grabiak*, 769 F.2d 729, 732-33, 226 USPQ 870, 872 (Fed. Cir. 1985). If the technology is unpredictable, it is less likely that structurally similar species will render a claimed species obvious because it may not be reasonable to infer that they would share similar properties. See, *e.g.*, *In re May*, 574 F.2d 1082, 1094, 197 USPQ 601, 611 (CCPA 1978) (*prima facie* obviousness of claimed analgesic compound based on structurally similar prior art isomer was rebutted with evidence demonstrating that analgesia and addiction properties could not be reliably predicted on the basis of chemical structure); *In re Schechter*, 205 F.2d 185, 191, 98 USPQ 144, 150 (CCPA 1953) (unpredictability in the insecticide field, with homologs, isomers and analogs of known effective insecticides having proven ineffective as insecticides, was considered as a factor weighing against a conclusion of obviousness of the claimed compounds). However, obviousness does not

require absolute predictability, only a reasonable expectation of success, *i.e.*, a reasonable expectation of obtaining similar properties. See, *e.g.*, *In re O'Farrell*, 853 F.2d 894, 903, 7 USPQ2d 1673, 1681 (Fed. Cir. 1988). But here, the PTO will no doubt agree that the technologic area – inhibition of HIV – is an unpredictable endeavor. This is yet another factor mitigating against obviousness.

The final office action challenges appellants' position on this point, but inexplicably confuses the issue of unpredictability with “surprising and unexpected results,” which are only required to rebut a *proper prima facie* case of obviousness. Where unpredictability undercuts any likelihood of success, as is the case here, there *is* no *prima facie* case. To the extent that this line of argument is instead meant to say that appellants have not provided evidence that treating HIV is unpredictable, appellants ask the Board to take judicial notice of this well-know and widely-accepted fact.

C. Summary

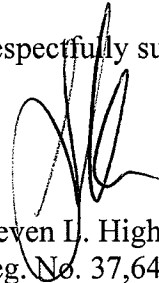
In sum, based on the foregoing factors and the evidence as a whole (*In re Bell*, 991 F.2d 781, 784, 26 USPQ2d 1529, 1531 (Fed. Cir. 1993); *In re Kulling*, 897 F.2d 1147, 1149, 14 USPQ2d 1056, 1057 (Fed. Cir. 1990)), the examiner has failed to make express fact-findings relating to the *Graham* factors and address the issues set forth above. The fact-findings should have specifically articulated what teachings or suggestions in the prior art would have motivated one of ordinary skill in the art to select the claimed species. *Kulling*, 897 F.2d at 1149, 14 USPQ2d at 1058; *Panduit Corp. v. Dennison Mfg. Co.*, 810 F.2d 1561, 1579 n.42, 1 USPQ2d 1593, 1606 n.42 (Fed. Cir. 1987). However, the final office action does no more than establish that the claimed species fall within the genus described by Lehrer, and does not address any of the factors set out above that would otherwise support the rejection. Thus, it is respectfully

submitted that the rejection is improper on its face and does not establish *prima facie* obviousness. Reversal of the rejection, based on the preceding, is respectfully requested.

D. Conclusion

In light of the foregoing, appellants respectfully submit that all claims are adequately described and non-obvious over the cited art. Therefore, reversal of all rejections is respectfully requested.

Respectfully submitted,

A handwritten signature in black ink, appearing to read 'S. Highlander', is written over the typed name and registration number.

Date: October 10, 2006

Steven L. Highlander
Reg. No. 37,642
Attorney for Appellants

VIII. CLAIMS APPENDIX

1. (Previously presented) A method for reducing the infectivity of an enveloped virus comprising contacting said virus with a first anti-viral peptide, said peptide comprising a chimeric theta defensin peptide selected from the group consisting of SEQ ID NO:31 and SEQ ID NO:32.
9. (Original) The method of claim 1, wherein the virus infects humans and is selected from the group consisting of HIV, HSV-1, HSV-2, EBV, varicella zoster virus, CMV, herpesvirus B, HHV6, HHV8, respiratory syncytial virus (RSV), influenza A, B and C viruses, hepatitis A, hepatitis B, hepatitis C, hepatitis G, smallpox, vaccinia virus, Marburg virus, ebola virus, dengue virus, West Nile virus, hantavirus, measles virus, mumps virus, rubella virus, rabies virus, yellow fever virus, Japanese encephalitis virus, Murray Valley encephalitis virus, Rocio virus, tick-borne encephalitis virus, St. Louis encephalitis virus, chikungunya virus, o'nyong-nyong virus, Ross River virus, Mayaro virus, human coronaviruses 229-E and OC43, vesicular stomatitis virus, sandfly fever virus, Rift Valley River virus, Lasa virus, lymphocytic choriomeningitis virus, Machupo virus, Junin virus, HTLV-I and -II.
18. (Original) The method of claim 1, further comprising contacting said virus with a second anti-viral agent.
19. (Original) The method of claim 18, wherein said second anti-viral agent is a second anti-viral peptide distinct from said first anti-viral peptide.
20. (Original) The method of claim 18, wherein said second anti-viral agent is non-peptide pharmaceutical agent.
21. (Original) The method of claim 20, wherein said non-peptide pharmaceutical agent is selected from the group consisting of a protease inhibitor, a nucleoside analog, a viral polymerase inhibitor, and a viral integrase inhibitor.

22. (Original) The method of claim 1, wherein said first anti-viral peptide is contacted with said virus at a concentration of about 0.1 to about 50 μg per ml.
23. (Original) The method of claim 22, wherein said first anti-viral peptide is contacted with said virus at a concentration of about 1 to about 25 μg per ml.
24. (Original) The method of claim 23, wherein said first anti-viral peptide is contacted with said virus at a concentration of about 3 to about 10 μg per ml.
27. (Original) The method of claim 1, wherein said virus is located in a living subject.
28. (Original) The method of claim 27, wherein said first anti-viral peptide is administered topically.
34. (Original) The method of claim 27, wherein said first anti-viral peptide is administered to a wound site.
35. (Original) The method of claim 27, wherein said patient is immunosuppressed.
36. (Original) The method of claim 27, wherein said subject is not infected with said virus, and first anti-viral peptide is administered prior to the virus contacting the subject.
37. (Original) The method of claim 27, wherein said first anti-viral peptide is administered subsequent to the virus contacting the subject.
38. (Original) The method of claim 37, wherein said subject is chronically infected with said virus.
40. (Original) The method of claim 37, wherein said subject is acutely infected with said virus.

IV. EVIDENCE APPENDIX

Exhibit 1 – Lehrer *et al.*, WO 02/085401

Exhibit 2 – James Darnell *et al.*, *Molecular Cell Biology* (3rd ed. 1998).

X. RELATED PROCEEDINGS APPENDIX

[NONE]

EXHIBIT 1

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International Bureau



(43) International Publication Date
31 October 2002 (31.10.2002)

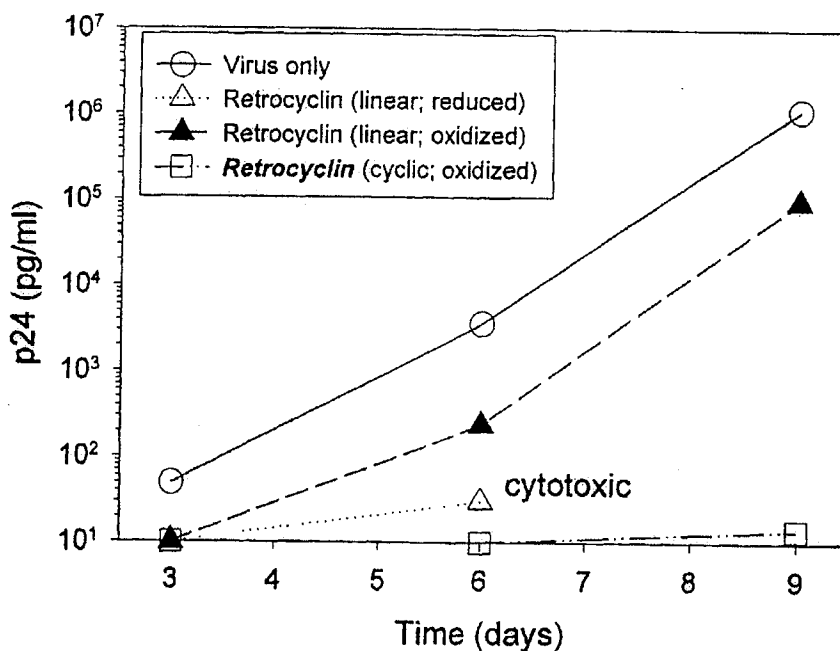
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- (71) Applicant (for all designated States except US): **THE REGENTS OF THE UNIVERSITY OF CALIFORNIA** [US/US]; 1111 Franklin Street, 5th Floor, Oakland, CA 94607 (US).
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- (74) Agent: **SHERWOOD, Pamela, J.**; Bozicevic, Field & Francis, L.L.P., 200 Middlefield Road, Suite 200, Menlo Park, CA 94025 (US).
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[Continued on next page]

(54) Title: RETROCYCLINS: ANTIVIRAL AND ANTIMICROBIAL PEPTIDES



(57) Abstract: Retrocyclin peptides are small antimicrobial agents with potent activity against bacteria and viruses. The peptides are nonhemolytic, and exhibit minimal *in vitro* cytotoxicity. A pharmaceutical composition comprising retrocyclin as an active agent is administered therapeutically to a patient suffering from a bacterial and/or viral infection, or to an individual facing exposure to a bacterial and/or viral infection, especially one caused by the HIV-1 retrovirus or other sexually-transmitted pathogens.

WO 02/085401 A1



Published:

- with international search report
- before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

RETROCYCLINS: ANTIVIRAL AND ANTIMICROBIAL PEPTIDES

STATEMENT AS TO FEDERALLY SPONSORED RESEARCH

This invention was made with government support under grant number AI22839 awarded
5 by the National Institutes of Health. The government has certain rights in the invention.

INTRODUCTION

Background

Natural polycationic antimicrobial peptides have been found in many different species of animals and insects and shown to have broad antimicrobial activity. In mammals, these
10 antimicrobial peptides are represented by two families, the defensins and the cathelicidins. Nearly all of these peptides have membrane affinity, and can permeate and permeabilize bacterial membranes, resulting in injury, lysis, and/or death to the microbes. In particular, the human peptides known as defensins are produced by mammalian and avian leukocytes (e.g. neutrophils, some macrophages) and epithelial cells.

15 Three defensin subfamilies exist in vertebrates: alpha-defensins, beta-defensins, and circular (theta) minidefensins. All derive from an ancestral gene that existed before reptiles and birds diverged, contain six cysteines, and have largely beta-sheet structures that are stabilized by three intramolecular disulfide bonds. RTD-1, a theta minidefensin, was recently detected in bone marrow from the rhesus monkey, *Macacca mulatta*. It had 18 residues and was circular,
20 having been formed by the fusion of two truncated alpha-defensin precursors ("demidefensins") each of which contributed 3 cysteines to the mature peptide. The cellular machinery responsible for processing these precursors remains operational in human leukocytes.

Alpha-defensins are largely beta sheet peptides that contain 29-35 amino acid residues, including 6 cysteines that form three intramolecular disulfide bonds. Because of the nature of the
25 cysteine pairings, the molecules are effectively macrocyclic. Four of these α -defensins, HNP 1-4, occur primarily in human neutrophils. HD-5 & 6 are found in Paneth cells, specialized cells of the small intestine's crypts. Human α -defensin genes contain three exons and two introns and are clustered on chromosome 8p23. They encode preprodefensins that contain ~100 residues which encode a signal peptide, polyanionic propiece and the C-terminal defensin domain. Mature
30 defensins are processed by sequential proteolysis.

Beta defensins are generally larger than α -defensins (35-40 residues) and may also be more ancient, since they occur in birds as well as mammals. Beta defensins are expressed in many different types of epithelial cells, and in some glands. In some cases, expression is constitutive; in others, it is inducible. Several β -defensin genes are located on 8 p23, adjacent to
35 the α -defensin genes- consistent with their common evolutionary ancestry. The disulfide pairing motif of beta defensins differs from that of α -defensins, however α and β -defensins have generally similar shapes.

The three-dimensional structure of many defensins comprises a complexly folded amphiphilic beta-sheet, with the polar face formed by its arginines and by the N- and C-terminal residues playing an important role in defining microbicidal potency and the antimicrobial spectrum. The antimicrobial effects of defensins are derived from their ability to permeabilize cell membranes and interact with viral envelopes, thereby exposing contents of the microorganism to the environment or abrogating viral infectivity. (See Gudmundsson *et al.* (1999) J Immunol Methods 232(1-2):45-54.) Antimicrobial peptides are reviewed by Hancock and Lehrer (1998) Trends in Biotechnology 16:82.

In general, the antiviral activities of antimicrobial peptides have not been extensively investigated. Although studies have reported that antimicrobial peptides, such as human neutrophil-derived defensins (α -defensins), are directly virucidal against herpes simplex virus (HSV), and adenovirus strains, only a few reports deal with anti-HIV-1 activity. T22 and T140, analogs of polyphemusins (peptides from horseshoe crabs), are active in inhibiting HIV-1 replication through binding to the chemokine receptor CXCR4. However, these peptides only inhibit the T cell-tropic (T-tropic; X4) strains that utilize CXCR4 as a coreceptor for entry and they are ineffective against strains that utilize CCR5 for entry (macrophage (M)-tropic "R5" viruses). Since sexual transmission is largely attributed to R5 infection, the potential of T22 and T140 as topical vaginal or rectal microbicides is limited.

One study indicated that protegrins (porcine-derived peptides) can inactivate HIV-1 virions. Another study showed that indolicidin, a 13 amino acid peptide isolated from bovine neutrophils, was reproducibly virucidal against HIV-1 only at very high concentrations (333 μ g/ml) of peptide. While the anti-HIV-1 activity of human α -defensins has not been reported, certain structural and functional similarities exist between the loop motifs of α -defensins and peptides derived from HIV-1 gp41 that may be required for viral fusion and infectivity.

Vaginal and rectal subepithelial stromal tissues are densely populated with dendritic cells (DC), macrophages and T-cells that express both CD4 and the HIV-1 coreceptors, CXCR4 and CCR5. Mechanisms whereby HIV-1 journeys across the mucosal epithelia are not clear, but may directly involve the epithelial cells. Once the virus reaches the lamina propria, it can either directly infect macrophages or T-cells or adhere to or infect DC whose traffic to the regional lymph nodes conveys them into sites of vigorous viral replication. A recent report suggests that binding of HIV-1 to DC is mediated by the C-type lectin DC-SIGN, independent of CD4 or chemokine receptors. Thus, mucosal factors which modulate steps in this process could affect the probability of transmission of HIV-1 infection.

There is a clinical need for novel antiviral and antimicrobial agents that have low toxicity against mammalian cells. The present invention addresses this need.

Relevant literature

Defensins are reviewed by Lehrer *et al.* (1992) *Ann. Rev. Immunol.* 11:105-128. Other endogenous antimicrobials are reviewed in Schonwetter *et al.* (1995) *Science* 267:1645-1648; Schroder (1999) *Cell Mol Life Sci.* 56:32-46 (1999); and Harwig *et al.* (1994) *FEBS Lett* 342:281-285.

5 Specific defensins are described in Tang *et al.* (1999) *Science* 286:498-502; Zimmermann *et al.* (1995) *Biochemistry* 34:13663-13671; Liu *et al.* (1997) *Genomics* 43:316-320; and Palfree & Shen (1994) GenBank U10267; Polley *et al.* GenBank AF238378 disclose the sequence of *Homo sapiens* chromosome 8p23 clone SCb-561b17.

Retrovirus infection and antiretroviral therapy are discussed in Wilson *et al.* (1995) *J. Infect. Dis.* 172:88-96; Wong *et al.* *Science* 278:1291-1295; and Yang *et al.* (1999) *J. Virol.* 73, 4582-4589.

SUMMARY OF THE INVENTION

Methods and compositions are provided for the use of retrocyclin peptides. Retrocyclin
15 peptides are small antimicrobial agents with potent activity against viruses, *e.g.* enveloped viruses such as retroviruses; and bacteria. These circular peptides are nonhemolytic and generally exhibit little or no *in vitro* cytotoxicity. Retrocyclins are equally effective against growing and stationary phase bacteria, and they retain activity against some bacteria in physiological, and high salt concentrations. Studies indicate that retrocyclins are also capable of conferring
20 immunity to human CD4⁺ cells against infection by HIV-1 *in vitro*. In addition, other circular mini-defensins also find use as anti-viral agents, particularly against human retroviruses.

A pharmaceutical composition comprising retrocyclin or other circular mini-defensins as an active agent is administered to a patient suffering from a viral infection. Alternatively, a pharmaceutical composition comprising retrocyclin or other circular mini-defensins or is
25 administered as a protective agent to a normal individual facing potential exposure to HIV viruses or pathogenic microbes. Retrocyclin is also effective at killing a variety of microbial organisms, *in vivo* and *in vitro*. Retrocyclin may be administered alone, or in combination with other bacteriocidal agents, *e.g.* antibiotics and/or other antiviral agents, and antiviral agents as a cocktail of effective peptides, etc. Retrocyclin-mediated killing is also useful for modeling and
30 screening novel antibiotics.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Circular minidefensins reduce HIV-1 infection of H9 cells. HIV-1 strain IIIB (MOI = 10⁻²) was incubated with 2.5 × 10⁵ H9 cells in the presence or absence of 20 µg/ml RTD-1,
35 RTD-2 or RTD-3. p24 antigen release was monitored by ELISA on days 3, 6, and 9. Assay sensitivity = 10 pg/ml.

Figure 2. Sequence comparison of human and rhesus demidefensins. The translated

sequences of rhesus demidefensin-1 mRNA and human retrocyclin mRNA are shown. Solid circles (●) indicate a stop codon in the corresponding cDNA. Vertical bars connect identical residues, and + signs connect similar residues. Residues represented in mature retrocyclin and RTD-molecules are boxed. The demidefensin-1 sequence (GenBank, AF184156) was derived
5 from the monkey mRNA (not shown).

Figure 3. Structural characterization of retrocyclin. (A) CD spectrum demonstrating the similarity in structure between retrocyclin and RTD-1, both at 0.5 mg/ml in a 1:1 mixture of trifluoroethanol in phosphate buffered saline at pH 7.4. (B) shows a hypothetical model of retrocyclin made by templating its sequence on the backbone of a similar peptide from porcine
10 neutrophils, Protegrin-1 (PDB accession code: 1PG1). (D) is a cartoon version of (B), wherein arginines are black, cysteines are grey and the other residues are identified by single letter code. (C) is a similar cartoon of rhesus RTD-1, indicating the similarity in structure with retrocyclin.

Figure 4. Effect of salt on antibacterial activity of circular minidefensins. Human retrocyclin and monkey RTD-1 were tested against our standard lab stains: *E. coli* ML-35p, *P. aeruginosa* MR 3007, *L. monocytogenes* EGD, and *S. aureus* 930918. The bars show MIC
15 values \pm SEM values that resulted from 3-6 radial diffusion assays per organism and assay condition.

Figure 5. Anti-HIV-1 activity of retrocyclin. Two strains of HIV-1 and two types of human target cells were used. The IIIB strain is T-cell tropic (X4) and utilizes the CXCR4 co-receptor for
20 entry; the JR-CSF strain is M-tropic (R5) and uses CCR5 for entry. PBMC signifies CD4⁺ selected peripheral blood mononuclear cells. Results indicate p24 antigen concentration in pg/ml, as determined by quantitative ELISA assay at Day 9 timepoint. (A) Two concentrations of retrocyclin (2 μ g/ml, 20 μ g/ml), 20 μ g/ml of the Rhesus circular defensin "RTD-1", and 20 μ g/ml of a horseshoe crab-derived peptide "T140", reported to only prevent X4 infections, were tested
25 in antiviral assays of against strain IIIB in H9 cells (n = 2-6 per peptide; error bars indicate SEM). (B) To confirm our results with primary human cells, similar assays were performed utilizing IIIB virus and CD4⁺ PBMC or (C) JR-CSF virus and CD4⁺ PBMC. Peptides were not cytotoxic at indicated concentrations, measured by trypan blue exclusion. Average of duplicate experiments are reported for studies with PBMCs. Assay sensitivity = 10 pg/ml.

30 Figure 6. Retrocyclin can inhibit HIV-1 spread when administered up to 24 hrs post-infection. Primary CD4⁺ PBMC were incubated with HIV-IIIB for 3 hours in the absence ("control", "t0", "t3", and "t24") or presence ("t0 only") of 20 μ g/ml retrocyclin. Cells were transferred to fresh R10-50 media that was either supplemented immediately with 20 μ g/ml retrocyclin ("t0"), or 3 or 24 hrs after transfer ("t3" and "t24", respectively). "Control" and "t0 only"
35 were not supplemented after transfer. p24 antigen was measured by ELISA as previously described.

Figure 7. Mature retrocyclin, but not premature forms, inhibit HIV-1 replication. H9 cells

were incubated with HIV-IIIB ($\text{MOI} = 10^{-2}$) for 3 hours in the absence or presence of $20\mu\text{g/ml}$ retrocyclin in three flavors: linear and reduced; linear and oxidized disulfide bonds; and the mature form (cyclic and oxidized). Assay sensitivity is 10 pg/ml .

Figure 8. Cytotoxicity of antimicrobial peptides against H9 cells. Retrocyclin, RTD-1 and PG-1 (a porcine-derived peptide with anti-HIV-1 activity) were tested for cytotoxicity using an MTT assay for cell proliferation. Note that the EC_{50} of Retrocyclin and RTD-1 were $>100\text{ }\mu\text{g/ml}$, concentrations well above their antiviral concentration.

Figures 9A and 9B are graphs depicting the activity of retrocyclin congeners against HIV-1 strains.

Figure 10 is a diagram depicting the structure of retrocyclin.

Figure 11 compares the antiretroviral activity of retrocyclin and RC-101 ($20\text{ }\mu\text{g/ml}$), by showing the p24 titers from day 9 CD4^+ PBMC (peripheral blood mononuclear cells) infected with HIV-1 strains IIIB or JR-CSF at the indicated MOI. RC-101 and retrocyclin were similarly effective in inhibiting HIV-1 replication at low MOI (A) and higher MOI (B).

Figure 12. Adding retrocyclin directly with HIV-1-IIIB does not reduce infection of H9 cells. Retrocyclin ($2\text{--}200\text{ }\mu\text{g/ml}$) was incubated with HIV-IIIB ($\text{MOI} = 10^{-2}$) diluted in R10 media prior to infecting H9 cells. p24 antigen release was measured by ELISA. Limit of detection = 10 pg/ml .

Figure 13. Retrocyclin and RC-101 inhibited the formation of HIV proviral DNA. Retrocyclin and RC-101 inhibited the formation of DNA from both early events (total HIV DNA) and later events (full-length HIV DNA) of reverse transcription. Data are an average of 2 experiments, except for RC-101 (1 experiment). "HI virus" is a heat-inactivated virus control for background levels of viral DNA.

Figure 14. Inactivation of HSV-1 and HSV-2 by retrocyclin and RC-101. Retrocyclin (left panel) and RC-101 (right panel) at the indicated concentrations were incubated with herpes simplex virus, type 1 (HSV-1) or HSV-2 for 2 hrs and then added to ME-180 cell monolayers. Cells were incubated at 37°C for 72 hrs, and cytotoxicity was measured with an MTT kit.

Figure 15 depicts sequences of human, ape and monkey retrocyclins.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Novel compositions and methods are provided for the use of retrocyclins and retrocyclin analogs as therapeutic and/or prophylactic agents. The peptides are effective at killing a variety of microbial organisms by direct microbicidal activity, and protect against viral infection by a virus
 5 by preventing viral uptake and/or blocking an early step in viral replication. Retrocyclin(s) are administered alone or in combination with other active agents to a patient suffering from an infection in a dose and for a period of time sufficient to reduce the patient population of pathogenic microbes or viruses. Alternatively, a pharmaceutical composition comprising retrocyclin or other circular mini-defensins or is administered as a protective agent to a normal
 10 individual facing potential exposure to HIV viruses or pathogenic microbes. In addition, other circular mini-defensins, including RTD-1, RTD-2 and RTD-3 and variants of retrocyclin find use as anti-viral agents.

Specific treatments of interest include, without limitation: using retrocyclin (*e.g.*, RC-101) or a retrocyclin analog to prevent or treat infection, for example by an enveloped virus, including
 15 enveloped retroviruses, more specifically by HIV-1, HIV-2 and related retroviruses that cause Acquired Immunodeficiency Syndrome (AIDS); aerosol administration to the lungs of patients with cystic fibrosis to combat infection or forestall the emergence of resistance to other inhaled antibiotics; instillation into the urinary bladder of patients with indwelling catheters to prevent
 infection; application to the skin of patients with serious burns; ophthalmic instillation, directly or in
 20 ophthalmic solutions, to treat or prevent infection; intravaginal application to treat bacterial vaginosis and/or prevent sexually transmitted disease such as HIV infection. The retrocyclins also may find use in the treatment of plant-pathogenic pseudomonads, in agricultural applications designed to prevent disease in and spoilage of food crops. The retrocyclins may be administered alone or in conjunction with other antiviral therapy.

25 The peptide form of retrocyclins provides a basis for further therapeutic development, by modification of the polypeptide structure to yield modified forms having altered biological and chemical properties. The native or modified forms are formulated in a physiologically acceptable carrier for therapeutic uses, or are otherwise used as an antimicrobial agent.

30

RETROCYCLIN COMPOSITIONS

For use in the subject methods, a naturally occurring or synthetic retrocyclin may be used. As used herein, retrocyclins are cyclic polypeptides comprising the amino acid sequence:

X₁ X₂ X₃ X₄ X₅ X₆ X₇ X₈ X₉ X₁₀ X₁₁ X₁₂ X₁₃ X₁₄ X₁₅ X₁₆ X₁₇ X₁₈
 wherein X₁ and X₁₈ are linked through a peptide bond,

disulfide crosslinks are formed between at least one of: X₃ and X₁₆; X₅ and X₁₄, and X₇
 35 and X₁₂, usually between at least two of such pairs, and preferably between the three pairs of amino acids, with the proviso that when such a crosslink is present, the crosslinked amino acids are both cysteines;

at least about three of amino acids X_1 to X_{18} are arginine or lysine, and the number of arginine or lysine residues may be four or more, five or more, or six or more. Preferred residues for arginine or lysine are X_4 , X_9 , X_{13} , and X_{18} ;

X_2 , X_5 , X_{11} , X_{15} are preferably aliphatic amino acids, e.g. isoleucine, leucine, valine, 5 phenylalanine, and alanine;

X_1 , X_8 , X_{10} and X_{17} are preferably glycine or alanine, usually glycine.

Retrocyclins are octadecapeptides that contain two linked nonapeptides that may be identical or different. A consensus nonapeptide has the sequence shown below, where the 10 bolded and underlined residues are invariant among the primate sequences identified herein. Substitutions found in the nonapeptide regions of other circular minidefensin precursors are shown below the consensus nonapeptide.

	Residue No	1 3 5 7 9
15	Consensus nonapeptide	<u>RCICGRGIC</u>
	Variant	L RL V
	Variant	T F
	Variant	V

From the consensus peptide and these variants, one can generate unique nonapeptide 20 sequences (herein termed n_1 , n_2 , etc.), Thus, n_1 could be linked to itself or any of the other nonapeptides ($n_1:n_1$, $n_1:n_2$, $n_1:n_3$, etc.), to generate unique octadecapeptides. To continue the process, n_2 could be linked to itself or to any other nonapeptide except n_1 , to generate additional unique octadecapeptides, and so forth.

The set of nonapeptides derived from these sequences (which are also provided in the 25 sequence listing as SEQ ID NO:19-64) is as follows:

	1 2 3 4 5 6 7 8 9	Mod'm*		1 2 3 4 5 6 7 8 9	Mod'n*
1	R C I C G R G I C	--	25	R C I C G L G V C	L6, V8
2	R C L C G R G I C	L3	26	R C L C R L G I C	L3, R5, L6
3	R C I C R R G I C	R5	27	R C L C R R G V C	L3, R5, V8
4	R C I C T R G I C	T5	28	R C L C R R G F C	L3, R5, F8
5	R C I C V R G I C	V5	29	R C L C T L G I C	L3, T5, L6
6	R C I C G L G I C	L6	30	R C L C T R G V C	L3, T5, V8
7	R C I C G R G V C	V8	31	R C L C T R G F C	L3, T5, F8
8	R C I C G R G F C	F8	32	R C L C V L G I C	L3, V5, L6
9	R C L C R R G V C	L3, R5	33	R C L C V R G V C	L3, V5, V8
10	R C L C T R G I C	L3, T5	34	R C I C G R G I C	L3, V5, F8
11	R C L C V R G I C	L3, V5	35	R C I C R L G V C	R5, L6, V8
12	R C L C G L G V C	L3, L6	36	R C I C R L G F C	R5, L6, F8
13	R C L C G R G V C	L3, V8	37	R C I C T L G V C	T5, L6, V8
14	R C L C G R G F C	L3, F8	38	R C I C T L G F C	T5, L6, F8
15	R C I C R R G V C	R5, V8	39	R C I C V L G V C	V5, L6, V8

16	R C I C R R G F C	R5, F8	40	R C I C V L G F C	V5, L6, F8
17	R C I C T R G V C	T5, V8	41	R C L C G L G V C	L3, R5, L6, V8
18	R C I C T R G F C	T5, F8	42	R C L C G L G I C	L3, R5, L6, F8
19	R C I C T L G I C	T5, L6	43	R C L C T L G V C	L3, T5, L6, V8
20	R C I C V L G F C	V5, L6	44	R C L C T L G I C	L3, T5, L6, F8
21	R C I C R L G I C	R5, L6	45	R C L C V L G V C	L3, V5, L6, V8
22	R C I C V R G V C	V5, V8	46	R C L C V L G I C	L3, V5, L6, F8
23	R C I C G R G F C	V5, F8	47		
24	R C I C G L G F C	L6, F8	48		

* residue modifications are shown in this column.

Retrocyclins of interest include cyclic peptides derived from the peptide sequence set forth in SEQ ID NO. 12, in particular a circular homodimer comprising a dimer of the amino acid sequence SEQ ID NO:12, aa 48-56. This retrocyclin has the structure (SEQ ID NO:1):

G I C R C I C G R G I C R C I C G R
X₁ X₂ X₃ X₄ X₅ X₆ X₇ X₈ X₉ X₁₀ X₁₁ X₁₂ X₁₃ X₁₄ X₁₅ X₁₆ X₁₇ X₁₈

5

Wherein X₁ and X₁₈ are joined by a peptide bond, X₂ and X₁₁, X₄ and X₉, and X₁₃ and X₁₈ are disulfide bonded.

Another retrocyclin of interest is the synthetic analog (SEQ ID NO:2)

G I C R C I C G K G I C R C I C G R
X₁ X₂ X₃ X₄ X₅ X₆ X₇ X₈ X₉ X₁₀ X₁₁ X₁₂ X₁₃ X₁₄ X₁₅ X₁₆ X₁₇ X₁₈

10 wherein X₁ and X₁₈ are joined by a peptide bond, X₂ and X₁₁, X₄ and X₉, and X₁₃ and X₁₈ are disulfide bonded. Other synthetic analogs, or congeners, or retrocyclin are set forth in SEQ ID NO:3-SEQ ID NO:10.

The sequence of the retrocyclin polypeptides may be altered in various ways known in the art to generate targeted changes in sequence. The polypeptide will usually be substantially
15 similar to the sequences provided herein, i.e. will differ by one amino acid, and may differ by two amino acids. The sequence changes may be substitutions, insertions or deletions.

The protein may be joined to a wide variety of other oligopeptides or proteins for a variety of purposes. By providing for expression of the subject peptides, various post-translational modifications may be achieved. For example, by employing the appropriate coding sequences,
20 one may provide farnesylation or prenylation. In this situation, the peptide will be bound to a lipid group at a terminus, so as to be able to be bound to a lipid membrane, such as a liposome.

Modifications of interest that do not alter primary sequence include chemical derivatization of polypeptides, e.g., acetylation, or carboxylation. Also included are modifications of glycosylation, e.g. those made by modifying the glycosylation patterns of a polypeptide during
25 its synthesis and processing or in further processing steps; e.g. by exposing the polypeptide to enzymes which affect glycosylation, such as mammalian glycosylating or deglycosylating enzymes. Also embraced are sequences that have phosphorylated amino acid residues, e.g.

phosphotyrosine, phosphoserine, or phosphothreonine.

Also included in the subject invention are polypeptides that have been modified using ordinary molecular biological techniques and synthetic chemistry so as to improve their resistance to proteolytic degradation or to optimize solubility properties or to render them more
5 suitable as a therapeutic agent. Analogs of such polypeptides include those containing residues other than naturally occurring L-amino acids, e.g. D-amino acids or non-naturally occurring synthetic amino acids.

The subject peptides may be prepared by *in vitro* synthesis, using conventional methods as known in the art. Various commercial synthetic apparatuses are available, for example,
10 automated synthesizers by Applied Biosystems, Inc., Foster City, CA, Beckman, *etc.* By using synthesizers, naturally occurring amino acids may be substituted with unnatural amino acids. The particular sequence and the manner of preparation will be determined by convenience, economics, purity required, and the like.

If desired, various groups may be introduced into the peptide during synthesis or during
15 expression, which allow for linking to other molecules or to a surface. Thus cysteines can be used to make thioethers, histidines for linking to a metal ion complex, carboxyl groups for forming amides or esters, amino groups for forming amides, and the like.

The polypeptides may also be isolated and purified in accordance with conventional methods of recombinant synthesis. A lysate may be prepared of the expression host and the
20 lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique. For the most part, the compositions which are used will comprise at least 20% by weight of the desired product, more usually at least about 75% by weight, preferably at least about 95% by weight, and for therapeutic purposes, usually at least about 99.5% by weight, in relation to contaminants related to the method of preparation of
25 the product and its purification. Usually, the percentages will be based upon total protein. Genetic sequences encoding demi-defensins are provided herein, e.g. SEQ ID NO:4, 7 and 9.

In one embodiment of the invention, the antimicrobial peptide consists essentially of a polypeptide sequence set forth in any one of SEQ ID NO:1-SEQ ID NO:10. By "consisting essentially of" in the context of a polypeptide described herein, it is meant that the polypeptide is
30 composed of the sequence set forth in the seqlist, which sequence may be flanked by one or more amino acid or other residues that do not materially affect the basic characteristic(s) of the polypeptide.

For some purposes of the invention, for example in the treatment and/or prevention of HIV infection, the active agent may be any one of the circular minidefensins, e.g. retrocyclin,
35 RTD-1, RTD-2 and RTD-3. Cyclic minidefensins resemble protegrins, antimicrobial β -sheet peptides. RTD-1 is derived from *Macacca mulatta*, and is a heterodimer containing tandem nonapeptide elements derived from the mature peptides set forth in SEQ ID NO:15 and SEQ ID

NO:17. RTD-2 is a homodimer containing, in tandem, two identical nonapeptide elements derived from the mature peptide set forth in SEQ ID NO:17. RTD-3 is a homodimer containing, in tandem, two identical nonapeptide elements derived from the mature peptide set forth in SEQ ID NO:15.

All three RTD's are circular molecules with 18 residues and three intramolecular disulfide bonds. Each RTD is formed by *in vivo* processing that trims and splices two precursor peptides ("demidefensins"), each of which contributes nine residues (including 3 cysteines) to the mature cyclic peptide. The 18 RTD-1 residues derive from two different demidefensin precursors, RTD-2 and -3 have tandem 9 residue repeats derived from a single demidefensin precursor.

10

RETROCYCLIN CODING SEQUENCES

The invention includes nucleic acids having a sequence set forth in SEQ ID NO:11; nucleic acids that hybridize under stringent conditions, particularly conditions of high stringency, to the sequence set forth in SEQ ID NO:11; genes corresponding to the provided nucleic acids; sequences encoding retrocyclins; and fragments and derivatives thereof. Other nucleic acid compositions contemplated by and within the scope of the present invention will be readily apparent to one of ordinary skill in the art when provided with the disclosure here. Genetic sequences of particular interest include primate sequences, *e.g.* human, chimpanzee, bonobo, orangutan, gorilla, *etc.*

Retrocyclin coding sequences can be generated by methods known in the art, *e.g.* by *in vitro* synthesis, recombinant methods, *etc.* to provide a coding sequence that corresponds to a linear retrocyclin polypeptide that could serve as an intermediate in the production of the cyclic retrocyclin molecule. Using the known genetic code, one can produce a suitable coding sequence. For example, the circular polypeptide of retrocyclin (SEQ ID NO: 1) is encoded by the sequence (SEQ ID NO:18) AGG TGC ATT TGC GGA AGA GGA ATT TGC AGG TGC ATT TGC GGA AGA GGA ATT TGC, but since the peptide is circular, it is somewhat arbitrary which codon is selected to be first, allowing this to be based on other criteria, *e.g.* relative efficiency in purification or cyclization of the predicted product. The polypeptide set forth in SEQ ID NO:2 is encoded by a similar sequence, wherein one of the arginine codons (AGA) is substituted with a lysine codon (AAA or AAG).

The nucleic acids of the invention include nucleic acids having sequence similarity or sequence identity to SEQ ID NO:11 or SEQ ID NO:18. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 10XSSC (0.9 M saline/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC. Sequence identity can be determined by hybridization under stringent conditions, for example, at 50°C or higher and 0.1XSSC (9 mM saline/0.9 mM sodium citrate). Hybridization methods and conditions are well known in the art, see, *e.g.*, U.S. patent no. 5,707,829. Nucleic acids that are substantially identical to the provided nucleic acid sequence, *e.g.* allelic variants,

genetically altered versions of the gene, *etc.*, bind to SEQ ID NO:11 or SEQ ID NO:18 under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes can be any species, *e.g.* primate species, particularly human; rodents, such as rats and mice; 5 canines, felines, bovines, ovines, equines, fish, yeast, nematodes, *etc.*

In one embodiment, hybridization is performed using at least 18 contiguous nucleotides (nt) of SEQ ID NO:1 and SEQ ID NO:18, or a DNA encoding the polypeptide of SEQ ID NO:1-10. Such a probe will preferentially hybridize with a nucleic acid comprising the complementary sequence, allowing the identification and retrieval of the nucleic acids that uniquely hybridize to 10 the selected probe. Probes of more than 18 nt can be used, *e.g.*, probes of from about 18 nt to about 25, 50, 100 or 250 nt, but 18 nt usually represents sufficient sequence for unique identification.

Nucleic acids of the invention also include naturally occurring variants of the nucleotide sequences (*e.g.*, degenerate variants, allelic variants, *etc.*). Variants of the nucleic acids of the 15 invention are identified by hybridization of putative variants with nucleotide sequences disclosed herein, preferably by hybridization under stringent conditions. For example, by using appropriate wash conditions, variants of the nucleic acids of the invention can be identified where the allelic variant exhibits at most about 25-30% base pair (bp) mismatches relative to the selected nucleic acid probe. In general, allelic variants contain 15-25% bp mismatches, and can contain as little 20 as even 5-15%, or 2-5%, or 1-2% bp mismatches, as well as a single bp mismatch.

The invention also encompasses homologs corresponding to the nucleic acids of SEQ ID NO:5, where the source of homologous genes can be any mammalian species, *e.g.*, primate species, particularly human; rodents, such as rats; canines, felines, bovines, ovines, equines, fish, yeast, nematodes, *etc.* Between mammalian species, *e.g.*, human and mouse, homologs 25 generally have substantial sequence similarity, *e.g.*, at least 75% sequence identity, usually at least 90%, more usually at least 95% between nucleotide sequences. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, *etc.* A reference sequence will usually be at least about 18 contiguous nt long, more usually at least about 30 nt long, and may extend to 30 the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as gapped BLAST, described in Altschul et al. Nucl. Acids Res. (1997) 25:3389-3402.

The subject nucleic acids can be cDNAs or genomic DNAs, as well as fragments thereof, particularly fragments that encode a biologically active polypeptide and/or are useful in the 35 methods disclosed herein. The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species

have contiguous exons, with the intervening introns, when present, being removed by nuclear RNA splicing, to create a continuous open reading frame encoding a polypeptide of the invention.

A genomic sequence of interest comprises the nucleic acid present between the initiation
5 codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally present in a native chromosome. It can further include the 3' and 5' untranslated regions found in the mature mRNA. It can further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, *etc.*, including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' and 3' end of the transcribed region.
10 The genomic DNA can be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' and 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for proper tissue, stage-specific, or disease-state specific expression.

The nucleic acid compositions of the subject invention can encode all or a part of the
15 subject polypeptides. Double or single stranded fragments can be obtained from the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, *etc.* Isolated nucleic acids and nucleic acid fragments of the invention comprise at least about 18, about 50, about 100, to about 200 contiguous nt selected from the nucleic acid sequence as shown in SEQ ID NO:11. For the
20 most part, fragments will be of at least 18 nt, usually at least 25 nt, and up to at least about 50 contiguous nt in length or more.

Probes specific to the nucleic acid of the invention can be generated using the nucleic acid sequence disclosed in SEQ ID NO:11, or a DNA encoding the polypeptide of SEQ ID NO:1-10. The probes are preferably at least about 18 nt, 25nt or more of the corresponding contiguous
25 sequence. The probes can be synthesized chemically or can be generated from longer nucleic acids using restriction enzymes. The probes can be labeled, for example, with a radioactive, biotinylated, or fluorescent tag. Preferably, probes are designed based upon an identifying sequence of one of the provided sequences. More preferably, probes are designed based on a contiguous sequence of one of the subject nucleic acids that remain unmasked following
30 application of a masking program for masking low complexity (*e.g.*, BLASTX) to the sequence, *i.e.*, one would select an unmasked region, as indicated by the nucleic acids outside the poly-n stretches of the masked sequence produced by the masking program.

The nucleic acids of the invention are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the nucleic acids, either as DNA or
35 RNA, will be obtained substantially free of other naturally-occurring nucleic acid sequences, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant," *e.g.*, flanked by one or more nucleotides with which it is not normally associated

on a naturally occurring chromosome.

Retrocyclin encoding nucleic acids can be provided as a linear molecule or within a circular molecule, and can be provided within autonomously replicating molecules (vectors) or within molecules without replication sequences. Expression of the nucleic acids can be regulated by their own or by other regulatory sequences known in the art. The nucleic acids of the invention can be introduced into suitable host cells using a variety of techniques available in the art, such as transferrin polycation-mediated DNA transfer, transfection with naked or encapsulated nucleic acids, liposome-mediated DNA transfer, intracellular transportation of DNA-coated latex beads, protoplast fusion, viral infection, electroporation, gene gun, calcium phosphate-mediated transfection, and the like.

Expression vectors may be used to introduce a retrocyclin coding sequence into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about one day, more usually for a period of at least about several days to several weeks. The gene or retrocyclin peptide may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of vesicles. Jet injection may also be used for intramuscular administration, as described by Furth *et al.* (1992) Anal. Biochem. **205**:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang *et al.* (1992) Nature **356**:152-154), where gold microprojectiles are coated with the stresscopin or DNA, then bombarded into skin cells.

METHODS OF USE

Formulations of retrocyclins are administered to a host suffering from an ongoing bacterial or viral infection or who faces exposure to a bacterial or viral infection. Antiviral compositions may also utilize other circular mini-defensins, e.g. RC-101, RTD-1, -2, and -3, alone or in combination with retrocyclin. Administration may be topical, localized or systemic, depending on the specific microorganism. Generally the dosage will be sufficient to decrease the microbial or viral population by at least about 50%, usually by at least 1 log, and may be by 2 or more logs. The compounds of the present invention are administered at a dosage that reduces the pathogen population while minimizing any side-effects. It is contemplated that the composition will be obtained and used under the guidance of a physician for in vivo use. Retrocyclins are particularly useful for killing *Listeria monocytogenes* and *Escherichia coli*, and

for preventing infection by certain viruses, particularly enveloped retroviruses, e.g. enveloped retroviruses such as HIV-1, HIV-2, FIV, and the like.

Retrocyclins are also useful for *in vitro* formulations to kill microbes, particularly where one does not wish to introduce quantities of conventional antibiotics. For example, retrocyclins may be added to animal and/or human food preparations, or to blood products intended for transfusion to reduce the risk of consequent bacterial or viral infection. This may be of particular interest since a common route of infection of *E. coli* and *L. monocytogenes* is the gastrointestinal tract. Retrocyclins may be included as an additive for *in vitro* cultures of cells, to prevent the overgrowth of microbes in tissue culture.

The susceptibility of a particular microbe or virus to killing or inhibition by retrocyclins may be determined by *in vitro* testing, as detailed in the experimental section. Typically a culture of the microbe is combined with retrocyclins at varying concentrations for a period of time sufficient to allow the protein to act, usually ranging from about one hour to one day. The viable microbes are then counted, and the level of killing determined. Two stage radial diffusion assay is a convenient alternative to determining the MIC or minimum inhibitory concentration of an antimicrobial agent.

Viral pathogens of interest include retroviral pathogens, e.g. HIV-1; HIV-2, HTLV, FIV, SIV, etc. Microbes of interest, but not limited to the following, include: *Citrobacter* sp.; *Enterobacter* sp.; *Escherichia* sp.; e.g. *E. coli*; *Klebsiella* sp.; *Morganella* sp.; *Proteus* sp.; *Providencia* sp.; *Salmonella* sp.; e.g. *S. typhi*, *S. typhimurium*; *Serratia* sp.; *Shigella* sp.; *Pseudomonas* sp., e.g. *P. aeruginosa*; *Yersinia* sp.; e.g. *Y. pestis*, *Y. pseudotuberculosis*, *Y. enterocolitica*; *Francisella* sp.; *Pasturella* sp.; *Vibrio* sp.; e.g. *V. cholerae*, *V. parahemolyticus*; *Campylobacter* sp., e.g. *C. jejuni*; *Haemophilus* sp., e.g. *H. influenzae*, *H. ducreyi*; *Bordetella* sp., e.g. *B. pertussis*, *B. bronchiseptica*, *B. parapertussis*; *Brucella* sp., *Neisseria* sp., e.g. *N. gonorrhoeae*, *N. meningitidis*, etc. Other bacteria of interest include *Legionella* sp., e.g. *L. pneumophila*; *Listeria* sp., e.g. *L. monocytogenes*; *Staphylococcus* sp., e.g. *S. aureus*; *Mycoplasma* sp., e.g. *M. hominis*, *M. pneumoniae*; *Mycobacterium* sp., e.g. *M. tuberculosis*, *M. leprae*; *Treponema* sp., e.g. *T. pallidum*; *Borrelia* sp., e.g. *B. burgdorferi*; *Leptospirae* sp.; *Rickettsia* sp., e.g. *R. rickettsii*, *R. typhi*; *Chlamydia* sp., e.g. *C. trachomatis*, *C. pneumoniae*, *C. psittaci*; *Helicobacter* sp., e.g. *H. pylori*, etc.

Various methods for administration may be employed. For the prevention of HIV infection, administration to mucosal surfaces is of particular interest, e.g. vaginal, rectal, etc. The polypeptide formulation may be given orally, or may be injected intravascularly, subcutaneously, peritoneally, by aerosol, ophthalmically, intra-bladder, topically, etc. For example, methods of administration by inhalation are well-known in the art. The dosage of the therapeutic formulation will vary widely, depending on the specific retrocyclin or demi-defensin to be administered, the nature of the disease, the frequency of administration, the manner of

administration, the clearance of the agent from the host, and the like. The initial dose may be larger, followed by smaller maintenance doses. The dose may be administered as infrequently as weekly or biweekly, or fractionated into smaller doses and administered once or several times daily, semi-weekly, *etc.* to maintain an effective dosage level. In many cases, oral administration
5 will require a higher dose than if administered intravenously. The amide bonds, as well as the amino and carboxy termini, may be modified for greater stability on oral administration.

Formulations

The compounds of this invention can be incorporated into a variety of formulations for
10 therapeutic administration. More particularly, the compounds of the present invention can be formulated into pharmaceutical compositions by combination with appropriate, pharmaceutically acceptable carriers or diluents, and may be formulated into preparations in solid, semi-solid, liquid or gaseous forms, such as tablets, capsules, powders, granules, ointments, solutions, suppositories, injections, inhalants, gels, microspheres, lotions, and aerosols. As such,
15 administration of the compounds can be achieved in various ways, including oral, vaginal, buccal, rectal, parenteral, intraperitoneal, intradermal, transdermal, intratracheal, *etc.*, administration. The retrocyclins may be systemic after administration or may be localized by the use of an implant or other formulation that acts to retain the active dose at the site of
implantation.
20 The compounds of the present invention can be administered alone, in combination with each other, or they can be used in combination with other known compounds (*e.g.*, perforin; anti-inflammatory agents, antibiotics, *etc.*) In pharmaceutical dosage forms, the compounds may be administered in the form of their pharmaceutically acceptable salts. The following methods and excipients are merely exemplary and are in no way limiting.

25 For oral preparations, the compounds can be used alone or in combination with appropriate additives to make tablets, powders, granules or capsules, for example, with conventional additives, such as lactose, mannitol, corn starch or potato starch; with binders, such as crystalline cellulose, cellulose derivatives, acacia, corn starch or gelatins; with disintegrators, such as corn starch, potato starch or sodium carboxymethylcellulose; with
30 lubricants, such as talc or magnesium stearate; and if desired, with diluents, buffering agents, moistening agents, preservatives and flavoring agents.

The compounds can be formulated into preparations for injections by dissolving, suspending or emulsifying them in an aqueous or nonaqueous solvent, such as vegetable or other similar oils, synthetic aliphatic acid glycerides, esters of higher aliphatic acids or propylene
35 glycol; and if desired, with conventional additives such as solubilizers, isotonic agents, suspending agents, emulsifying agents, stabilizers and preservatives.

The compounds can be utilized in aerosol formulation to be administered via inhalation.

The compounds of the present invention can be formulated into pressurized acceptable propellants such as dichlorodifluoromethane, propane, nitrogen and the like.

The compounds can be used as lotions, for example to prevent infection of burns, by formulation with conventional additives such as solubilizers, isotonic agents, suspending agents, 5 emulsifying agents, stabilizers and preservatives.

Furthermore, the compounds can be made into suppositories by mixing with a variety of bases such as emulsifying bases or water-soluble bases. The compounds of the present invention can be administered rectally via a suppository. The suppository can include vehicles such as cocoa butter, carbowaxes and polyethylene glycols, which melt at body temperature, yet 10 are solidified at room temperature.

Unit dosage forms for oral, vaginal or rectal administration such as syrups, elixirs, and suspensions may be provided wherein each dosage unit, for example, teaspoonful, tablespoonful, tablet or suppository, contains a predetermined amount of the composition containing one or more compounds of the present invention. Similarly, unit dosage forms for 15 injection or intravenous administration may comprise the compound of the present invention in a composition as a solution in sterile water, normal saline or another pharmaceutically acceptable carrier.

Implants for sustained release formulations are well-known in the art. Implants are formulated as microspheres, slabs, etc. with biodegradable or non-biodegradable polymers. For 20 example, polymers of lactic acid and/or glycolic acid form an erodible polymer that is well-tolerated by the host. The implant containing retrocyclins is placed in proximity to the site of infection, so that the local concentration of active agent is increased relative to the rest of the body.

The term "unit dosage form", as used herein, refers to physically discrete units suitable as 25 unitary dosages for human and animal subjects, each unit containing a predetermined quantity of compounds of the present invention calculated in an amount sufficient to produce the desired effect in association with a pharmaceutically acceptable diluent, carrier or vehicle. The specifications for the unit dosage forms of the present invention depend on the particular compound employed and the effect to be achieved, and the pharmacodynamics associated with 30 the compound in the host.

The pharmaceutically acceptable excipients, such as vehicles, adjuvants, carriers or diluents, are readily available to the public. Moreover, pharmaceutically acceptable auxiliary substances, such as pH adjusting and buffering agents, tonicity adjusting agents, stabilizers, wetting agents and the like, are readily available to the public.

35 Typical dosages for systemic administration range from 0.1 μ g to 100 milligrams per kg weight of subject per administration. A typical dosage may be one tablet taken from two to six times daily, or one time-release capsule or tablet taken once a day and containing a

proportionally higher content of active ingredient. The time-release effect may be obtained by capsule materials that dissolve at different pH values, by capsules that release slowly by osmotic pressure, or by any other known means of controlled release.

Those of skill will readily appreciate that dose levels can vary as a function of the specific compound, the severity of the symptoms and the susceptibility of the subject to side effects. Some of the specific compounds are more potent than others. Preferred dosages for a given compound are readily determinable by those of skill in the art by a variety of means. A preferred means is to measure the physiological potency of a given compound.

The use of liposomes as a delivery vehicle is one method of interest. The liposomes fuse with the cells of the target site and deliver the contents of the lumen intracellularly. The liposomes are maintained in contact with the cells for sufficient time for fusion, using various means to maintain contact, such as isolation, binding agents, and the like. In one aspect of the invention, liposomes are designed to be aerosolized for pulmonary administration. Liposomes may be prepared with purified proteins or peptides that mediate fusion of membranes, such as Sendai virus or influenza virus, etc. The lipids may be any useful combination of known liposome forming lipids; including cationic or zwitterionic lipids, such as phosphatidylcholine. The remaining lipid will be normally be neutral or acidic lipids, such as cholesterol, phosphatidyl serine, phosphatidyl glycerol, and the like.

For preparing the liposomes, the procedure described by Kato *et al.* (1991) J. Biol. Chem. 266:3361 may be used. Briefly, the lipids and lumen composition containing peptides are combined in an appropriate aqueous medium, conveniently a saline medium where the total solids will be in the range of about 1-10 weight percent. After intense agitation for short periods of time, from about 5-60 sec., the tube is placed in a warm water bath, from about 25-40° C and this cycle repeated from about 5-10 times. The composition is then sonicated for a convenient period of time, generally from about 1-10 sec. and may be further agitated by vortexing. The volume is then expanded by adding aqueous medium, generally increasing the volume by about from 1-2 fold, followed by shaking and cooling. This method allows for the incorporation into the lumen of high molecular weight molecules.

30

Formulations with Other Active Agents

For use in the subject methods, retrocyclins may be formulated with other pharmaceutically active agents, particularly other antimicrobial agents. Other agents of interest include a wide variety of antibiotics, as known in the art. Classes of antibiotics include penicillins, e.g. penicillin G, penicillin V, methicillin, oxacillin, carbenicillin, nafcillin, ampicillin, etc.; penicillins in combination with β -lactamase inhibitors, cephalosporins, e.g. cefaclor, cefazolin, cefuroxime, moxalactam, etc.; carbapenems; monobactams; aminoglycosides; tetracyclines; macrolides; lincomycins; polymyxins; sulfonamides; quinolones; cloramphenical; metronidazole;

spectinomycin; trimethoprim; vancomycin; *etc.*

Cytokines may also be included in a retrocyclin formulation, *e.g.* interferon γ , tumor necrosis factor α , interleukin 12, *etc.*

Antiviral agents, *e.g.* acyclovir, gancyclovir, *etc.*, and other circular mini-defensins (theta defensins) may also be included in retrocyclin formulations.

EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (*e.g.* amounts, temperature, concentrations, *etc.*) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

15

Sexual and mother-to-neonate (vertical) transmission through mucosal surfaces have been the most common routes of HIV-1 spread throughout the world. Although much attention has been focused on vaccine development for HIV-1, progress has been slow and there is an urgent need to find alternative approaches to prevent infections caused by HIV-1. Self-applied prophylactic agents to prevent mucosal, particularly vaginal or rectal, transmission of HIV-1 have the advantage of empowering vulnerable receptive partners to take effective measures for their own protection. In a search for novel compounds active against HIV-1, it was discovered that certain antimicrobial peptides, the circular minidefensins from the rhesus macaque, could inhibit HIV-1 replication. This prompted an investigation as to whether humans produce circular minidefensins. Although there is no evidence that these proteins are produced in humans, clearly some primate ancestors once made retrocyclin, because it continues to exist in contemporary humans as an expressed pseudogene.

After discovering an mRNA molecule in human bone marrow that was highly homologous to rhesus circular minidefensins (88.9% identity at the nucleotide level), solid phase peptide synthesis was used to create the peptide ("retrocyclin") whose sequence it encoded. Retrocyclin belongs to the θ defensin subfamily (also referred to as cyclic minidefensins). The antimicrobial properties of retrocyclin resemble those of rhesus θ -defensins. However, retrocyclin is highly effective in preventing the infection of CD4⁺ cells by X4 and R5 strains of HIV-1 *in vitro*.

35

Example 1

Circular minidefensins can block HIV-1 replication

It is shown herein that retrocyclin, a circular minidefensin, is potently active against both

X4 and R5 strains of HIV-1. The initial descriptions of circular minidefensins came from studies of *Macaca mulatta*, the rhesus macaque monkey. The first such peptide, RTD-1, was called a rhesus theta defensin (RTD), which are also referred to as "cyclic minidefensins". The peptides encoded by the mRNA precursors may be referred to as "demidefensins".

5 RTD-2 and RTD-3, which was isolated from the bone marrow of rhesus monkeys, are circular, 18 amino acid peptides that contained three intramolecular disulfide bonds. They are similar to RTD-1, the circular (θ) defensin previously described by Tang *et al.* However, whereas the 18 residues of RTD-1 represent spliced 9 amino acid fragments derived from two different minidefensin precursors, RTD-2 and -3 comprise tandem 9 residue repeats derived from a single
10 RTD-1 precursor. Thus, circular minidefensins are processed by a novel post-translational system that can generate a degree of effector molecule diversity without requiring commensurate genome expansion.

Retrocyclin and the other circular minidefensins we prepared were synthesized, folded, circularized and purified essentially. The antiviral activities of RTD-1, RTD-2 and RTD-3 are shown
15 in **Figure 1**. For these studies, the X4 HIV-1 strain IIIB was utilized.

Immortalized CD4⁺.H9 cells, which are permissive for infection with this strain, were maintained in RPMI supplemented with 10% heat-inactivated fetal calf serum (FCS), 10 mM HEPES, 2 mM glutamine, 100 U of penicillin/ml, and 10 μ g of streptomycin/ml (R10 media). Cells (2.5×10^5 /100 μ l) were incubated with virus (multiplicity of infection (MOI) = 10^{-2}) in the
20 presence or absence of 20 μ g/ml RTD-1, RTD-2 or RTD-3 for 3 hrs at 37 °C/5% CO₂. The cells were washed in R10 media, seeded in 48-well tissue culture plates in 1 ml R10 media, and incubated at 37 °C/5% CO₂ for 9 days. Aliquots of cell supernatant were removed at the specified time points and analyzed by a sensitive ELISA (DuPont NEN) that quantitates p24 antigen of HIV-1. The three circular rhesus minidefensins were similarly active, inhibiting HIV-1
25 by 100-1000 fold by 9 days post-inoculation (note the logarithmic scale).

Example 2

Identification and structural characterization of retrocyclin.

To search for human circular minidefensins, two primers were prepared based on the
30 monkey minidefensin cDNA sequences (GenBank AF 184156, 184157, 184158). When PCR was performed on Marathon-Ready human bone marrow cDNA (Clontech, Palo Alto, CA), a \approx 264 bp amplified product was recovered. To obtain its 3' and 5' side sequences, Marathon-Ready human bone marrow cDNA was amplified using a 3'-RACE kit (Gibco BRL, Gaithersburg, MD) and 5'-RACE kit from Boehringer Mannheim (Indianapolis, IN).

35 At the nucleotide level, this product (retrocyclin) was \sim 89% identical to the demidefensin precursors of rhesus RTD-1 (called precursors 1a and 1b). Figure 2 shows the peptide sequences of demidefensin 1 and preproretrocyclin. Residues incorporated into the mature

circular minidefensins are boxed and all stop codons are represented by solid circles. Although a stop codon within the human transcript's signal sequence should abort translation, the otherwise high conservation of rhesus and human mRNA's suggested that humans may have acquired this mutation relatively recently in primate evolution.

5 Three orangutan retrocyclin genes have been sequenced. One of these clones has the silencing stop codon in the signal sequence and therefore resembles human retrocyclin. The other two orangutan genes appear to be functional, *i.e.* when translated they would produce demi-defensins, the precursors of cyclic minidefensins.

Human leukocytes were examined for the presence of retrocyclin or similar peptides, but, 10 as expected from the presence of the signal sequence's stop codon, none was found. Thus synthetic retrocyclin represents the circular minidefensin that would have formed: a) if the signal sequence mutation were absent, and b) if the precursor underwent homologous pairing so that its boxed residues (see Figure 2) formed both halves of the circular molecule.

In phylogenetic studies of the retrocyclin demidefensin gene, the premature stop codon in 15 the signal sequence was found to be present in four anthropoid species (humans, gorillas, chimpanzees, pygmy chimpanzees) and not present in the genes of a fifth (orangutans). The demidefensin gene also appears intact (*i.e.*, no premature stop codon) in the two catarrhine (Old World Monkey) species examined to date, *Macaca mulatta* and *Macaca nemestrina*. These findings suggest that native retrocyclin peptides were last produced by a primate ancestor of 20 humans and other anthropoids that lived between 6 and 15 million years ago (mya). This is between the time that orangutan and human lineage diverged (15 mya) and before the divergence of the chimpanzee and human lineages (6 mya). These ongoing studies of primate phylogeny may yield sequence information about additional cyclic minidefensins whose native counterparts are extinct.

25 **Retrocyclin synthesis.** Peptides were synthesized at a 0.25 mmol scale with a Perkin-Elmer ABI 431A Synthesizer, using pre-derivatized polyethylene glycol polystyrene arginine resin (PerSeptive Biosystems, Framingham, MA), FastMoc™ chemistry, and double coupling for all residues. The crude peptide was reduced under nitrogen, for 15 hours at 50 °C with excess dithiothreitol in 6 M guanidine.HCl, 0.2 M Tris.HCl and 0.2 mM EDTA (pH 8.2). The reaction was 30 stopped with glacial acetic acid (final concentration, 5%) and the reduced peptide was stored under nitrogen until purified by RP-HPLC. After this step, the peptide appeared homogeneous and its mass (1942.5, by MALDI-TOF MS) agreed well with its theoretical mass. The reduced peptide (0.1 mg/ml) was oxidized, cyclized and purified essentially as described by Tang *et al.*, *supra*. The MALDI-TOF MS mass of retrocyclin (1918.5 Da) agreed well with its expected mass. 35 CD spectra were obtained at 25 °C from an AVIV 62DS spectropolarimeter (AVIV, Lakewood, NJ).

RTD-1 and retrocyclin have very similar CD spectra, with largely β -sheet structures

stabilized by disulfide linkages and connected by turns (Figure 3A). Antimicrobial peptides with similar spectra include tachyplesins, protegrins, and circularized defensins. Figure 3B, a backbone ribbon model of retrocyclin, was made by templating its sequence on the structure of protegrin PG-1 and cyclizing it. The resulting structure was annealed by molecular dynamics
 5 and energy minimized. Figure 3D is a cartoon version of Figure 3B, designed primarily to show the placement of the cysteine and arginine molecules. Figure 3C is a similar cartoon of rhesus RTD-1.

Retrocyclin is a selectively salt-insensitive antibacterial peptide. The effects of NaCl on
 10 the antimicrobial activity of retrocyclin and RTD-1, from two-stage radial diffusion assays, are compared in Figure 4. The peptides showed very similar behavior. Under low salt conditions, both peptides were highly effective (minimal inhibitory concentration (MIC) < 3 µg/ml) against all four test organisms: *Pseudomonas aeruginosa*, *Escherichia coli*, *Listeria monocytogenes* and *Staphylococcus aureus*. Their strong activity against *E. coli* and *L. monocytogenes* persisted in
 15 physiological (100 mM) NaCl, and even hypersalinity (175 mM NaCl) was only modestly inhibitory. In contrast, neither peptide was effective (MIC > 50 µg/ml) against *S. aureus* or *P. aeruginosa* in physiological or high salt concentrations. Retrocyclin's activity is likely to be
 preserved in the ionic concentration of the vaginal mucosa.

Retrocyclin potently inhibits HIV-1 replication of R5 and X4 viruses. The antiretroviral
 20 properties of retrocyclin are shown in Figure 5. Either HIV-1-permissive H9 cells were used as targets, or primary CD4⁺ lymphocytes from HIV-1-seronegative donors generated from freshly purified peripheral blood mononuclear cells (PBMC) stimulated with a CD3-CD8 bispecific monoclonal antibody. After approximately 7 days, when 98% of these cells co-expressed CD3 and
 25 CD4, they were infected with HIV-1 with or without retrocyclin or other test peptide. These cells were maintained in RPMI containing 10% FCS supplemented with 2 mM glutamine, 100 U of penicillin/ml, 10 µg of streptomycin/ml, and 50 U of interleukin 2/ml (R10-50 media).

Retrocyclin (10-20 µg/ml) afforded complete suppression of viral replication to CD4⁺-selected PBMC challenged with two different strains of HIV-1: IIIB (an X4 strain) and JR-CSF (an
 30 R5 strain), or H9 human T cells challenged with IIIB. Note that the concentration of p24 antigen is presented on a *log-scale* and that the rhesus circular minidefensins, RTD-1 (Figure 5 and Figure 1) and RTD-2 and RTD-3 were protective to a lesser extent than retrocyclin. Additionally, the antiretroviral activities of T140 (20 µg/ml; Figure 5) and T22, analogs of polyphemusins from horseshoe crabs that were previously shown to protect against X4, but not R5, infections, were
 35 confirmed in the present study. Microbicides, such as retrocyclin, that target both X4 and R5 viruses be more effective than agents that preferentially inhibit viruses of a single tropism.

Examining the effect of adding retrocyclin at various times pre- and post-HIV-1 infection.

To determine if retrocyclin is effective against HIV-1 when added post-infection, we either: 1) added retrocyclin at the time of HIV-1 infection, then washed away the peptide, or 2) added retrocyclin at various times post-infection. Primary CD4⁺ PBMC were incubated with HIV-III_B (Figure 6) or HIV-JR-CSF for 3 hours in the presence or absence of 20 µg/ml retrocyclin. The cells were subsequently washed in media, and incubated an additional 9 days. Retrocyclin (20 µg/ml) was added back to some of the cultures at time points specified in Figure 6 and infection was monitored by p24 ELISA as previously described. Although retrocyclin was most active when administered at the time of infection, and when present in culture throughout the 9 day incubation, retrocyclin administered as late as 24 hours after initial infection still reduced the p24 concentration by nearly 1000-fold.

Cyclization and oxidation are necessary for retrocyclin's antiviral activity.

Mature retrocyclin was prepared by a three-step process. Its two intermediate forms, as well as the final retrocyclin product were tested in our standard assay of HIV-1 infectivity: p24 ELISA of HIV-III_B infection of H9 cells (Figure 7). Intermediate 1 (open triangles) is the linearized retrocyclin octadecapeptide with 6 reduced cysteine thiol groups. Intermediate 2 (closed triangles) is a noncyclic b-hairpin octadecapeptide with 3 intramolecular cystine disulfide bonds. Retrocyclin (open squares), is a cyclized octadecapeptide with 3 disulfide bonds. Note that only the "mature" form of retrocyclin was antiviral, compared to control (no retrocyclin; open circles). Unlike retrocyclin, the linearized octadecapeptide was highly cytotoxic, as measured by trypan blue exclusion, and treated cells did not survive past 6 days.

Retrocyclin is not cytotoxic.

Cytotoxicity determinations were made with a Cell Proliferation Kit from Boehringer Mannheim used according to the manufacturer's instructions. The procedure measures the reduction of the yellowish MTT molecule (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide) to a dark blue formazan. Retrocyclin exhibited little to no cytotoxicity against H9 cells (Figure 8) and ME-180 cervical carcinoma cells at 100 µg/ml, a concentration that is far higher than the concentrations required for complete protection against HIV-1 infection (10 µg/ml). Additionally, neither 20 µg/ml retrocyclin nor RC-101 were cytotoxic to HIV-1-infected H9 cells and CD4⁺ PBMC as measured by trypan-blue exclusion (Table 1). Retrocyclin was not hemolytic for human erythrocytes.

Table 1. Cytotoxicity of 20 µg/ml peptide against H9 cells and CD4⁺ PBMC (peripheral blood mononuclear cells) as measured by Trypan blue exclusion.

Cells; virus	no peptide*	Retrocyclin
CD4 ⁺ PBMC; no virus	1.07	0.98

H9; IIIB	0.78	1.20
CD4 ⁺ PBMC; IIIB	1.71	1.68
CD4 ⁺ PBMC; JR-CSF	0.90	1.58

* Values expressed as the average number of cells $\times 10^5$ /ml for 2-3 experiments. N.D. = no data.

Construction and characterization of retrocyclin congeners. To date, we have constructed over a dozen congeners of retrocyclin, "RC-101", "RC-102", "RC-103", etc. and have used them to commence a structure-activity analysis of the retrocyclin's antiviral and antimicrobial effects. These peptides, whose sequences are shown in Table 2, were synthesized, oxidized, cyclized, and purified as described above for retrocyclin. RC-101 was prepared because retrocyclin, a circular peptide without free N-terminal or side-chain amine groups, is not well suited for fluorescent-conjugation. RC-101 is identical in sequence to retrocyclin (RC-100) except for the presence of an Arg₉→Lys₉ substitution. This modification preserves the net cationic charge of the peptide and provides an available epsilon-amino group in lysine's side chain. Importantly, RC-101 was as active as retrocyclin in protecting cells from infection by HIV-1 (Figure 11), indicating that substitutions in the primary sequence of retrocyclins can be made without losing anti-retroviral activity. The labeling of RC-101 with amine-reactive probes will be described in a later section.

Five additional analogues (RC 110-114) have been synthesized, cyclized and purified. RC-110 (Inverso-*enantioretrocyclin*), a cyclic peptide composed exclusively of D amino acids, has a sequence that is identical to retrocyclin, but with its residues placed in reverse order.

The ability of RC100 and several analogues described in Table 2 to protect cells from infection by X4 (HIV-IIIB) and R5 (JR-CSF) strains of HIV-1 is shown in Figure 9. The structure of retrocyclin itself is shown in Figure 10, with its residues numbered to correspond to Table 2.

The p24 assay results shown in Figure 9 are on a logarithmic scale. A horizontal reference line that passes through 10^0 on the ordinate scale corresponds to 1 pg/ml of p24 antigen. Results from 3 experiments (each performed with PBMC from a different donor) are shown. Retrocyclin was uniformly protective against both strains of HIV-1 in all of the experiments. Most of the mono-tyrosine substituted amino acid congeners of retrocyclin (RC-102, RC-103, RC-105, RC-106, and RC-108) were either inactive or only modestly active in inhibiting HIV-1 infection by Strain IIIB. In contrast, RC-102, RC-103 and RC-104 showed considerable ability to protect cells from infection by the JR-CSF strain (R5).

These results allow some hypotheses about the mechanism of action of retrocyclins to be formulated. Because RC-112 (*enantioretrocyclin*) was relatively ineffective, chiral interactions between retrocyclin and one or more receptors on the cell and/or virus surface are likely to participate in the protective mechanism. Since certain analogues (RC-102, RC-103 and perhaps RC-104) manifested substantial activity against the R5 strain but were relatively ineffective

against the X4 strain, the mechanisms whereby retrocyclin inhibits these strains are not identical.

The lack of efficacy of RC-106, RC-107 and RC-108 (each containing a tyrosine for arginine replacement) suggests that ionic interactions involving the positively charged arginine residues in position 4,9, and 13 of retrocyclin (see the model in Figure 10) with oppositely charged groups (e.g., phosphate) on the surface of the target cell or HIV-1 virion also participate in the process. In preliminary surface plasmon resonance (SPR) experiments, we have observed that retrocyclin binds with high affinity to certain sphingolipids (e.g., galactosylceramide) that are present in cell-surface rafts, and have been implicated in the cellular uptake of HIV-1 virions.

Table 2. Primary amino acid sequence of selected retrocyclin congeners.

SEQ ID NO:	Peptide	Name (or comment)	Avg. MW (Da)	Amino acid sequence
1	RC-100*	Retrocyclin	1918.4	GICRCICGRGICRCICGR
2	RC-101	R ₉ K-Retrocyclin	1890.4	GICRCICGKGICRCICGR
3	RC-102	I ₆ Y-Retrocyclin	1968.5	GICRCYCGRGICRCICGR
4	RC-103	I ₁₅ Y-Retrocyclin	1968.5	GICRCICGRGICRCYCGR
5	RC-104	I ₂ Y-Retrocyclin	1968.5	GYCRCICGRGICRCICGR
6	RC-105	I ₁₁ Y-Retrocyclin	1968.5	GICRCICGRGYCRCICGR
7	RC-106	R ₄ Y-Retrocyclin	1925.4	GICYCICGRGICRCICGR
8	RC-107	R ₉ Y-Retrocyclin	1925.4	GICICICGYGICRCICGR
9	RC-108	R ₁₃ Y-Retrocyclin	1925.4	GICICICGRGICYCICGR
10	RC-109**			GICICICGRGICRCICGY
19	RC-110	Inverso-enantio-Retrocyclin	1918.4	RGICRCICGRGICRCICG (ALL D)
20	RC-111	Inverso-retrocyclin	1918.4	RGICRCICGRGICRCICG
21	RC-112	enantio-retrocyclin	1918.4	GICRCICGRGICRCICGR (all D)
22	RC-113	enantio-RC-101	1890.4	GICRCICGKGICRCICGR (all D)
23	RC-114	RC-101/103 hybrid	1940.4	GICRCICGKGICRCYCGR

With the exception of RC-109, all of the above peptides are cyclic. * RC-100 is a synonym for retrocyclin, RC-111 (inverso-retrocyclin) is composed of L-amino acids; RC-110, 112 and 113 are composed exclusively of D-amino acids. RC-109 failed to cyclize, and has not been tested further.

15

Retrocyclin does not directly inactivate HIV-1. To determine if retrocyclin directly inactivated HIV-1 virions, HIV-IIIB (MOI 10⁻²) was incubated with 2 µg/ml, 20 µg/ml, or 200 µg/ml retrocyclin for 30 min at room temperature in R10 media. The mixture was diluted 190-fold in R10 media, to dilute retrocyclin below its effective antiviral concentrations (no significant antiviral activity at <2 µg/ml; n = 5), and used to infect 5 × 10⁵ H9 CD4⁺ cells. Viral replication was measured by collecting supernatant for 9 days at 3 day intervals to quantify HIV-1 p24 antigen by ELISA (Figure 12). HIV titer was not reduced with the highest concentration (200 µg/ml) of retrocyclin, demonstrating that retrocyclin does not target the virion directly. In this respect, the actions of retrocyclin are different from the direct inactivation of herpes simplex virus previously

observed with human and rabbit α -defensins.

Retrocyclin binds to T1 cells. Since retrocyclin does not directly inactivate HIV-1 virions, the ability of retrocyclin to interact with a cellular target was determined, using RC-101, a 5 Arg₉→Lys₉ congener of retrocyclin that retained the antiretroviral activity of the parent molecule. RC-101 was conjugated to the amine-reactive fluorescent dye, BODIPY-FL (Molecular Probes), according to the manufacturer's protocol. The conjugate (RC-101_{BODIPY-FL}) was purified by reverse-phase HPLC and resuspended in 0.01% acetic acid at up to 240 μ g/ml. RC-101_{BODIPY-FL} (20 μ g/ml) was incubated with 2.5×10^5 CD4⁺-selected PBMC cells for 15 min at room 10 temperature, washed once in fresh R10-50 media. Specimens were imaged on a Leica TCS-SP Confocal Microscope (Heidelberg, Germany) equipped with an argon laser for excitation of BODIPY-FL and phycoerythrin (PE). Images were collected with Leica Confocal Software. RC-101_{BODIPY-FL} bound to the cell membrane, mostly in patches. Patching ("microaggregation") has been reported to occur with hormone-occupied epidermal growth factor receptors (95); and 15 "rafts" are involved in signaling through the confinement of chemokine receptors to discrete regions of the cell membrane. RC-101_{BODIPY-FL} colocalizes with phycoerythrin (PE)-labeled monoclonal antibodies directed against CXCR4, CCR5 and CD4, but does not with PE-labeled isotype control antibodies. Thus, retrocyclin aggregates in the same "rafts" as the receptor and coreceptors for HIV-1. In addition, RC-101_{BODIPY-FL} aggregated in patches where CD4, CXCR4 20 and CCR5 levels were weak or absent.

A flow cytometry experiment was performed to examine binding of BODIPY-labeled RC-101. T1 cells were incubated for 1 hr at 37 °C \pm 20 μ g/ml RC-101_{BODIPY-FL}, washed with R10 media, and fixed in 2% paraformaldehyde/PBS. Cells were analyzed by fluorescence-activated cell sorting (FACS) on a Becton-Dickinson FACScan. Live cells (10^4 events) were gated and analyzed by 25 CellQuest. Two peaks were present, which may represent non-specific and specific cellular binding.

Retrocyclin inhibits HIV replication at an early step (reverse transcription or before). To determine whether retrocyclin blocked the formation of proviral DNA in HIV-JR-CSF-inoculated 30 CD4⁺-selected PBMC, quantitative real time PCR was performed. This method is more sensitive than measuring p24 release and can detect infection even when p24 values may be affected by virus carried over from the original inoculum. CD4⁺-selected PBMC (10^6 cells) were incubated in 250 μ l at 37 °C/5% CO₂ for 3 hr with either HIV-1 strain JR-CSF (MOI = 0.1), heat-inactivated virus (background control), JR-CSF + 20 μ g/ml retrocyclin or JR-CSF + 20 μ g RC-101. Cells 35 were washed and resuspended in 1 ml R10-50 media, and incubated for an additional 9 hours. Following incubation, cells were pelleted at 300 \times g, removed of overlying supernatant, and stored at -80 °C until analyzed by real time PCR. Retrocyclin and RC-101 inhibited the formation

of HIV-1 proviral DNA (Figure 13), indicating that retrocyclin acts early, either to inhibit reverse transcription or preceding events.

Some retrocyclins are slightly active against herpes simplex virus (HSV). To determine if the activity of retrocyclin against HIV-1 was specific or representative of a more global antiviral effect, it was tested its ability to prevent HSV infection *in vitro*. A quantitative microplate assay was used to screen retrocyclin and retrocyclin-congeners for their ability to inactivate HSV type 1 (HSV-1) and HSV-2. The assay utilized small amounts of peptide and simultaneously evaluated for peptide-induced cytotoxicity. In brief, final peptide concentrations of 2-50 $\mu\text{g/ml}$ were incubated with virus stocks for 2 hrs and added directly to ME-180 human cervical carcinoma cells. Cultures were incubated at 37°C for 72 hrs and cytotoxicity was measured using a 3-(4,5-dimethylthiazon-2-yl)-2,5-diphenyltetrazolium bromide (MTT) cell proliferation kit (Boehringer-Mannheim, Germany). Calculations to assess antiviral activity and compute "percent protection" are delineated in (90). Retrocyclin afforded modest protection against HSV-2, but not HSV-1 (Figure 14). In contrast, RC-102 and RC-103 were less antiviral than retrocyclin. However, RC-101 was modestly protective against HSV-1 and nearly completely protective against HSV-2. The substitution of Arg₉→Lys₉ produced a retrocyclin congener that retained activity against HIV-1, bacteria and fungi, and was more active against HSV.

Example 3

Sequences of Retrocyclins

Three orangutan clones that represent at east two different retrocyclin genes. The sequences are shown in Figure 15. The stop codons in orangutan clone 19 are identical to those in human retrocyclin. Accordingly, clone 19 also represents an expressed pseudogene. Overall, 132/143 (92.3%) of translated products (including stop codons) from orangutan clone 19 and the human retrocyclin gene are identical. The translation products of orangutan clones 20 and 21 are identical in 141/143 (98.6%) sites. Both clones lack a silencing stop codon in their signal sequence, and should be capable of producing a functional demidefensin whose tandem nonapeptide elements (underlined) would produce a peptide identical to human retrocyclin. The predicted translation products of orangutan clone 20 and human retrocyclin are identical in 129/143 (90.2%) of positions. All three orangutan clones, #19, 20 and 21 came from the DNA of a single orangutan. It remains to be determined if the genes they represent are alleles, or if the retrocyclin locus has undergone duplication and additional retrocyclin genes remain to be found.

As shown in Figure 15, this portion of the human retrocyclin gene encodes four stop codons (●). The first of these occurs near the end of the putative signal sequence and should abort translation. The second stop codon occurs after cysteine 3, and marks the end of the putative demidefensin sequence. The third stop codon comes after the CCR residues and

marks the customary termination of an α -defensin. The final stop codon occurs after the FES tripeptide in a non-expressed region of the gene.

One Gorilla retrocyclin clone has been sequenced. Its translation product is identical to human retrocyclin in 139/143 (97.2%). The sequence is shown in an alignment with the human
5 sequence in Figure 15. The silencing stop codon (●) is present in the signal sequence. Consequently, this clone represents an expressed pseudogene.

Note that the chimp (*Pan troglodytes*) and the Bonobo (*Pan paniscus*) genes contain the first stop codon (●) in the signal sequence, but both lack the retrocyclin-generating stop codon after cysteine 3 in the defensin-region. From these features, the chimp would appear to have
10 silenced an α -defensin gene. There is an additional mutation (cysteine to glycine) in the chimp's nonapeptide region (double underlined), which was presumably acquired after the gene had been silenced by the signal sequence mutation)

Unlike human retrocyclin, the pigtail and rhesus macaque genes lack a stop codon (●) in their signal sequences. Both macaque genes have acquired a stop codon in a nontranslated
15 portion of their gene, between cysteines 4 and 5 of the original defensin domain.

All publications and patent applications cited in this specification are herein incorporated
by reference as if each individual publication or patent application were specifically and
individually indicated to be incorporated by reference. The citation of any publication is for its
20 disclosure prior to the filing date and should not be construed as an admission that the present
invention is not entitled to antedate such publication by virtue of prior invention.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and
25 modifications may be made thereto without departing from the spirit or scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated retrocyclin peptide.
2. The isolated retrocyclin peptide according to Claim 1, wherein said peptide is
5 encoded by one or more primate genetic sequences.
3. The isolated retrocyclin peptide according to Claim 2, wherein said peptide is encoded by one or more human genetic sequences.
- 10 4. The isolated retrocyclin peptide according to Claim 1, wherein said peptide is linear.
5. The isolated retrocyclin peptide according to Claim 1, wherein said peptide is circular.
- 15 6. The isolated retrocyclin according to Claim 1, wherein said peptide is two linked nonapeptides, wherein each nonapeptide sequence is independently selected from the group consisting of SEQ ID NO:19 to SEQ ID NO:64.
- 20 7. A cyclic polypeptide comprising the amino acid sequence:
X₁ X₂ X₃ X₄ X₅ X₆ X₇ X₈ X₉ X₁₀ X₁₁ X₁₂ X₁₃ X₁₄ X₁₅ X₁₆ X₁₇ X₁₈
wherein X₁ and X₁₈ are linked through a peptide bond,
disulfide crosslinks are formed between at least one of: X₃ and X₁₆; X₅ and X₁₄; and X₇
and X₁₂ with the proviso that when such a crosslink is present, the crosslinked amino acids are
both cysteines;
25 at least about three of amino acids X₁ to X₁₈ are arginine or lysine;
X₂, X₅, X₁₁, X₁₅ are aliphatic amino acids; and
X₁, X₈, X₁₀ and X₁₇ are glycine or alanine.
8. The cyclic polypeptide according to Claim 7, wherein three pairs of cysteines are
30 crosslinked.
9. The cyclic polypeptide of Claim 7, wherein said polypeptide comprises the amino acid sequence set forth in SEQ ID NO:1-10.
- 35 10. The cyclic polypeptide of Claim 7, wherein said polypeptide comprises the amino acid sequence set forth in SEQ ID NO:1, 2, 5 or 8.

11. The cyclic polypeptide of Claim 1, and a pharmaceutically acceptable excipient.

12. An isolated nucleic acid encoding a primate retrocyclin or a peptide set forth in SEQ ID NO:1-10.

5

13. An isolated nucleic acid according to Claim 12, wherein said retrocyclin is human.

14. The isolated nucleic acid according to Claim 13, wherein said nucleic acid comprises the sequence set forth in SEQ ID NO:11.

10

15. A method for preventing retroviral infection in a cell, the method comprising: administering an effective dose of a circular minidefensin or retrocyclin to said cell.

16. The method according to Claim 14, wherein said cell is of a type that is susceptible to bacterial or viral infection.

17. A method for killing microbial organisms, the method comprising: administering an effective dose of retrocyclin to said microbial organisms.

18. A method for administering retrocyclin as a therapeutic agent to a patient with an established microbial or viral infection.

19. A method for administering retrocyclin as a prophylactic agent to prevent a microbial or viral infection in a patient at risk of developing such infection.

25

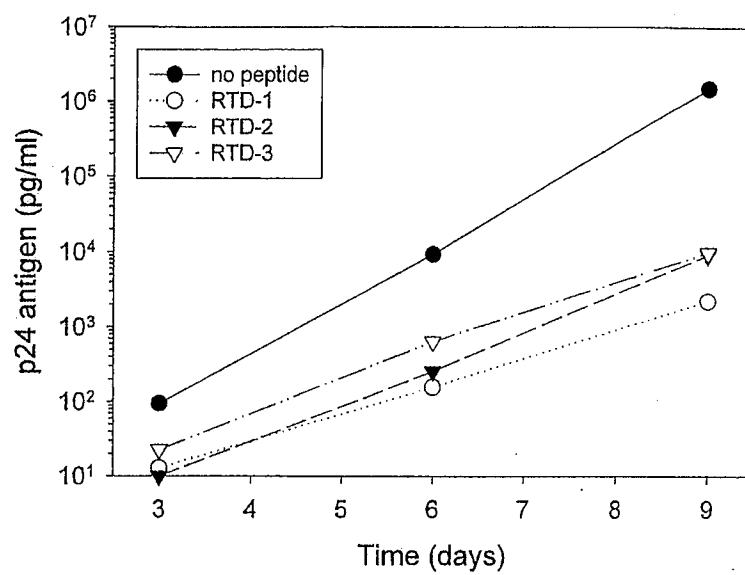


FIGURE 1

FIGURE 2

Demidefensin-1	MRTFALLTAMLLLVALLHAQAEARQARADEAAAAQQPGADDQGMHSFTRPENAAL	55
	+	
Human Retrocyclin	MRTFALLTAMLLLVALL•AQAEPQARADEAAAAQEPPGADDQEMAHFTWHESAAL	55
Demidefensin-1	PLSESARGLRCLCRRGVCQLL•RRIGSCAFRG•LCRICCR•	96
	+ +	
Human Retrocyclin	PLSDSARGLRCTCGRGICRL•RRFGSCAFRGTLHRICCR•	96

FIGURE 3

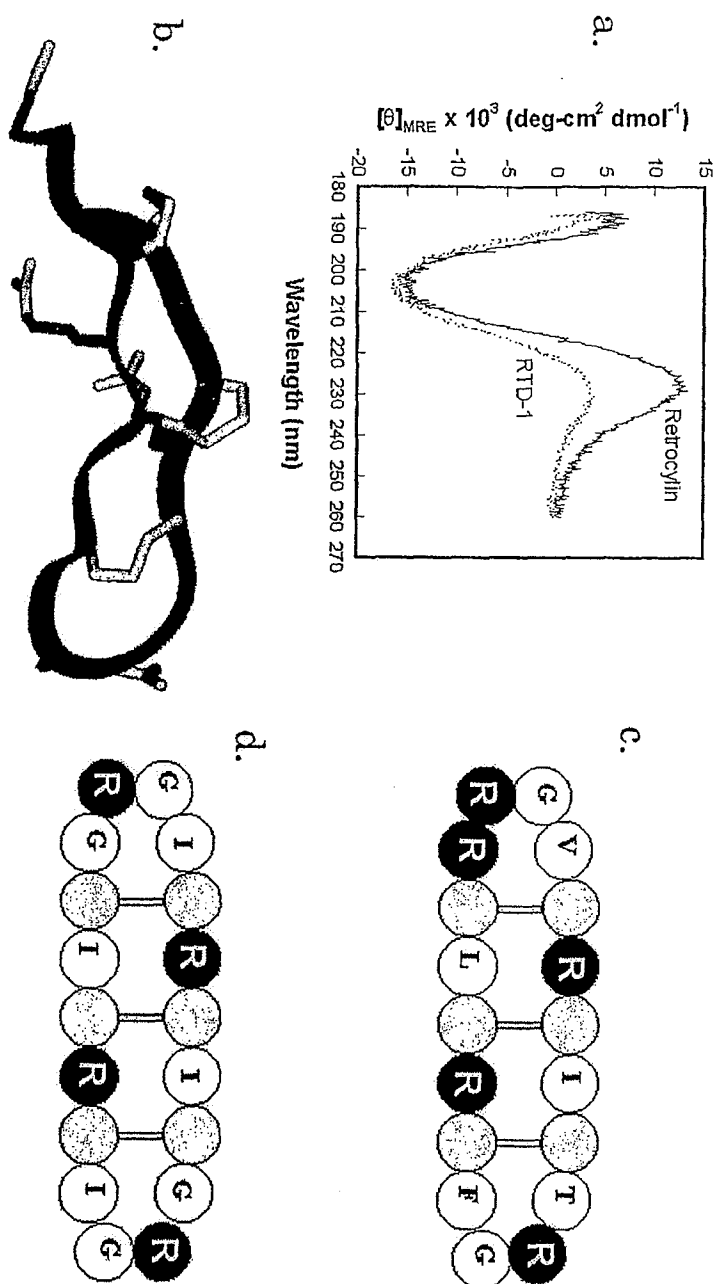


FIGURE 4

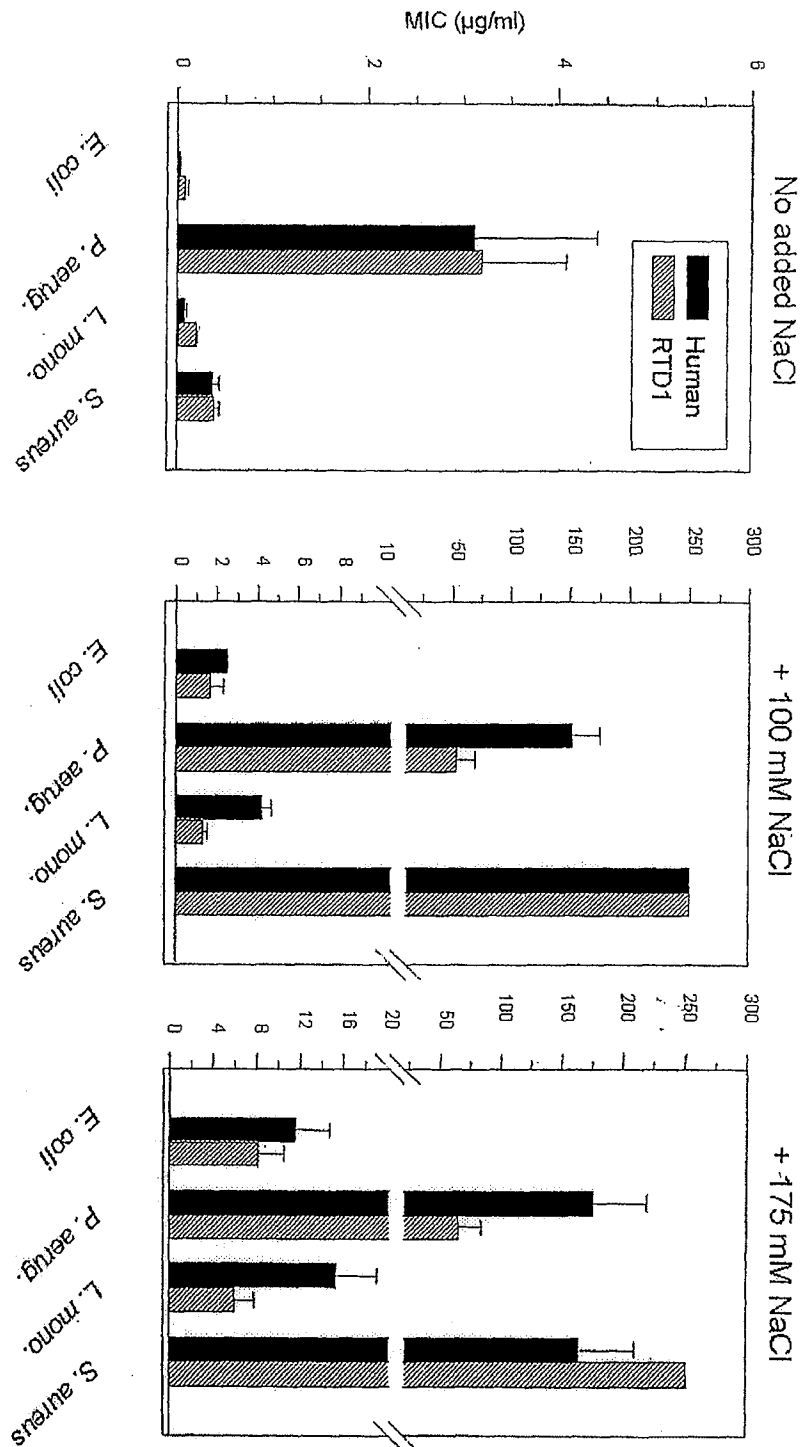


FIGURE 5

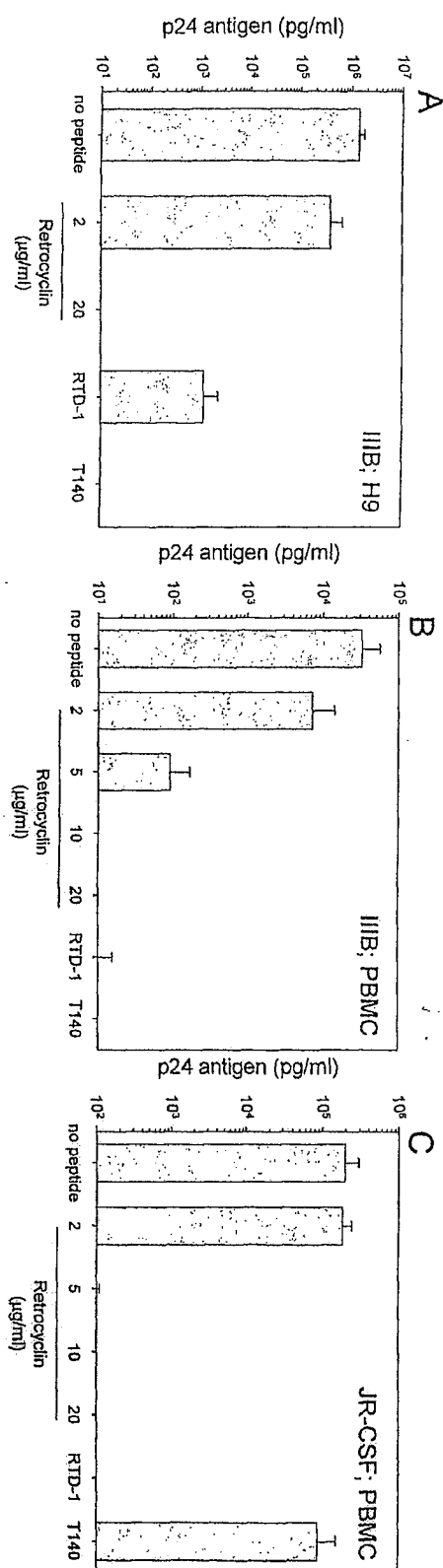


FIGURE 6

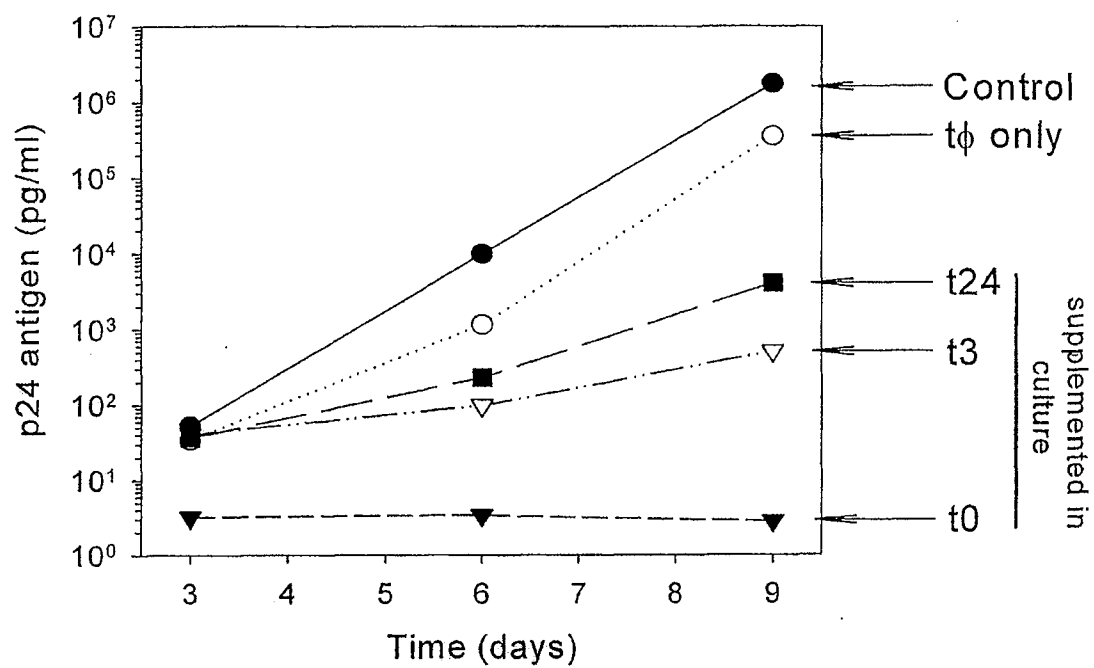


FIGURE 7

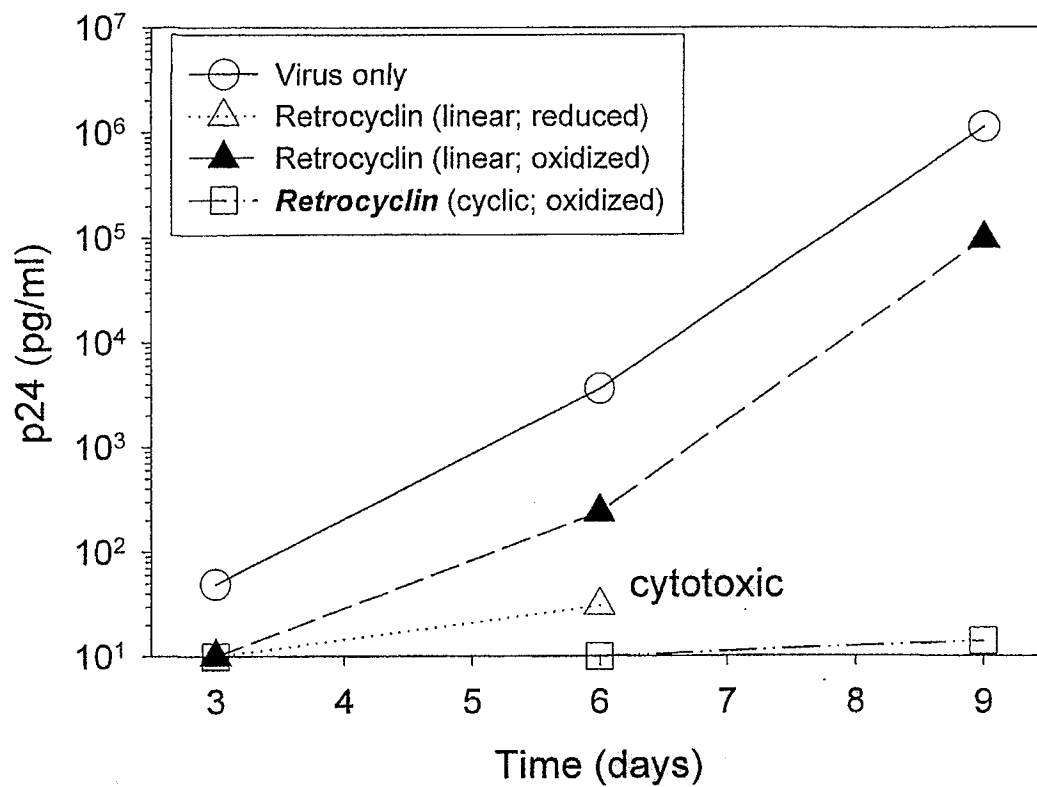


FIGURE 8

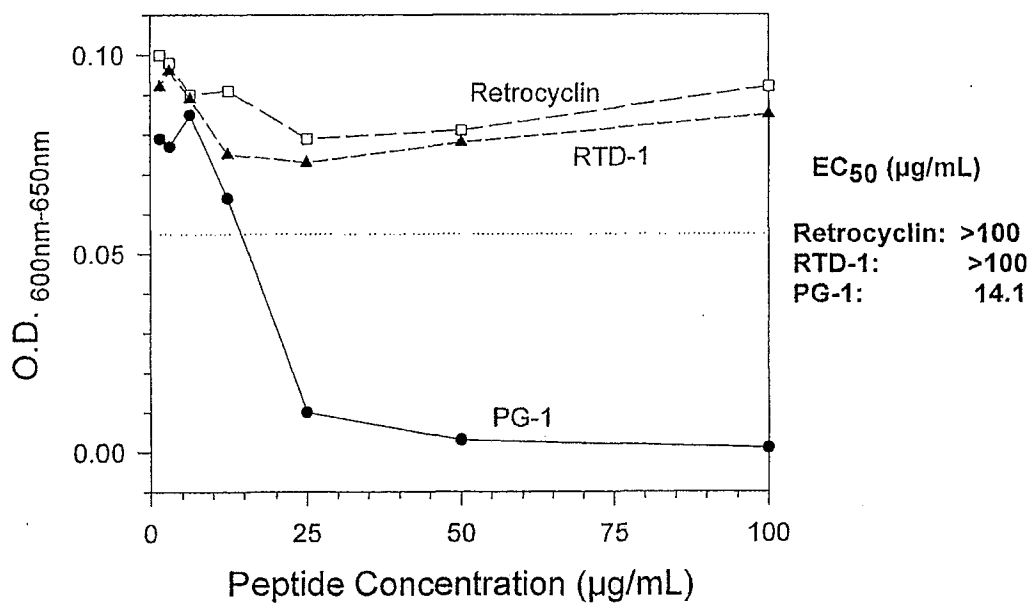


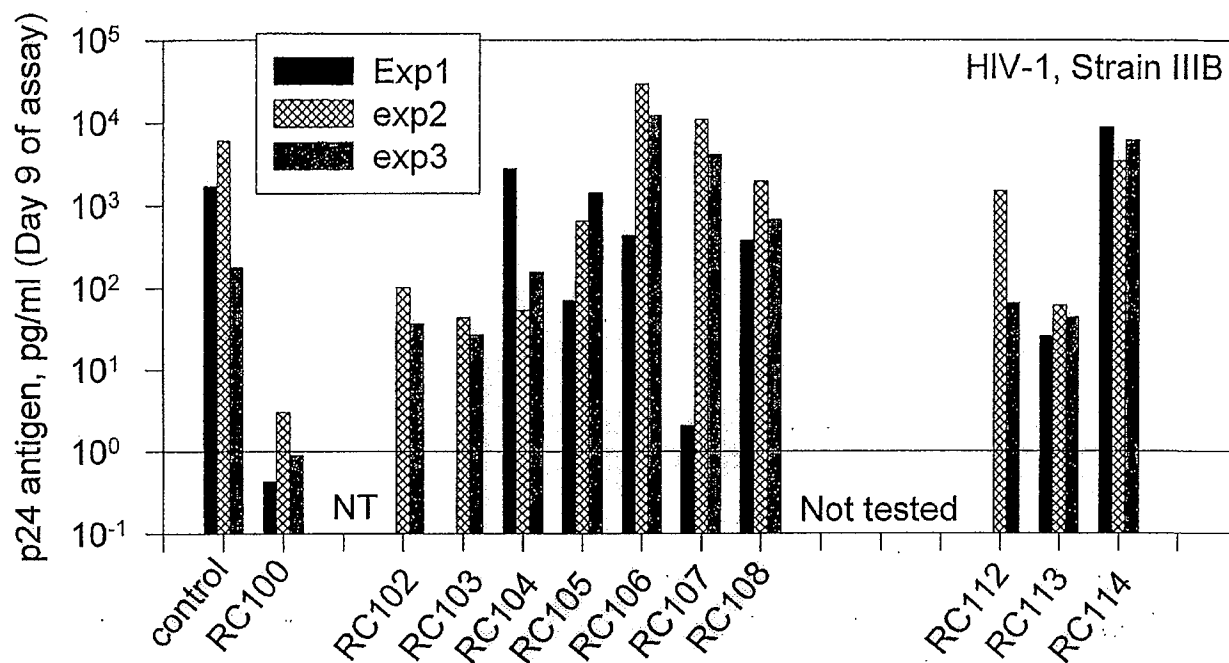
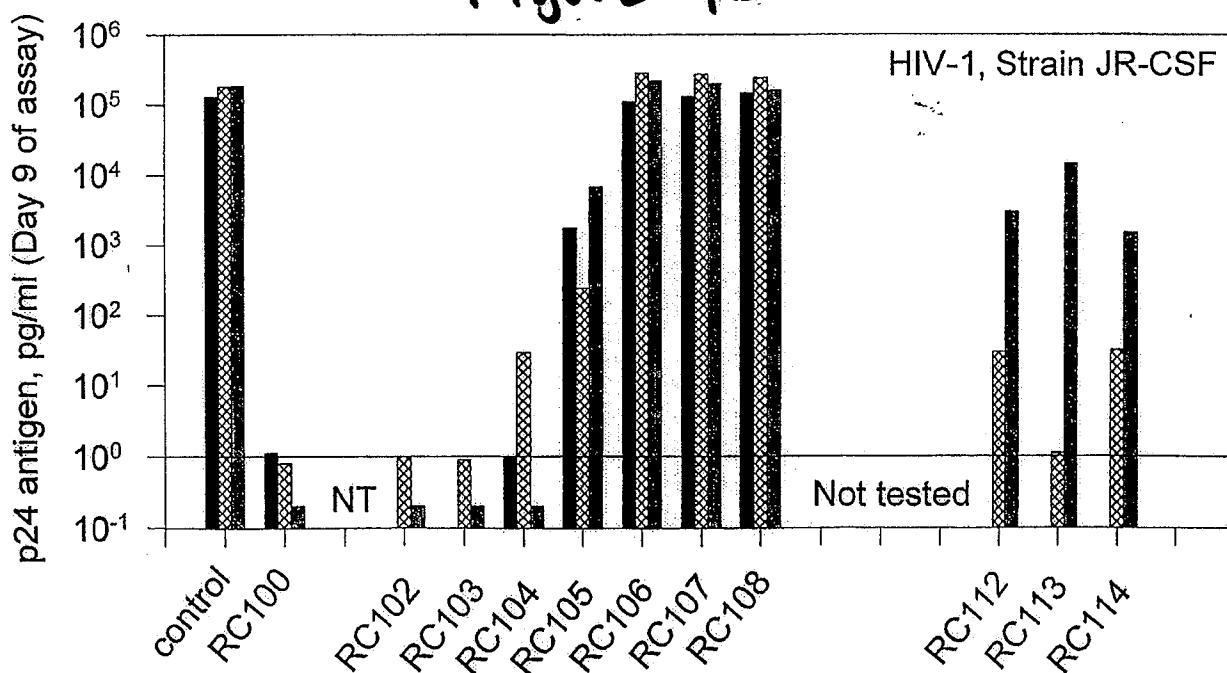
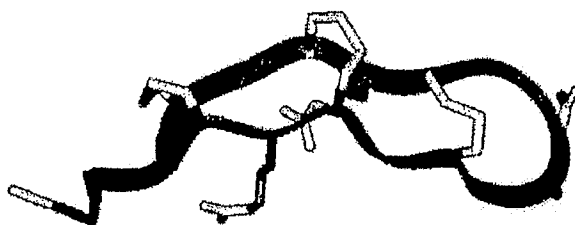
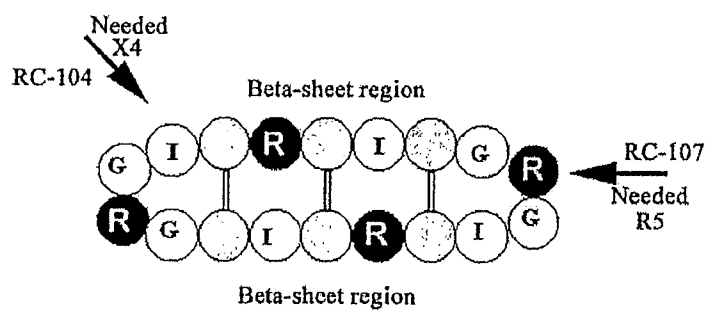
Figure 9 A**Figure 9B**

FIGURE 10



RETROCYCLIN

FIGURE 11

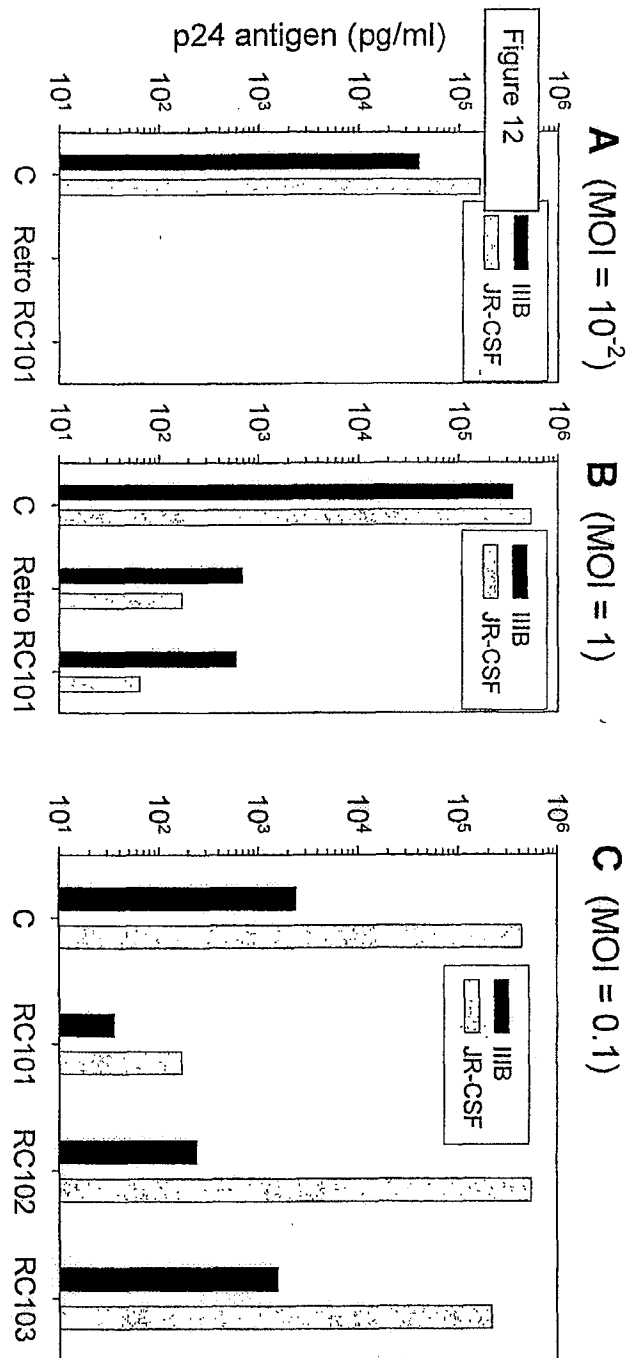


FIGURE 12

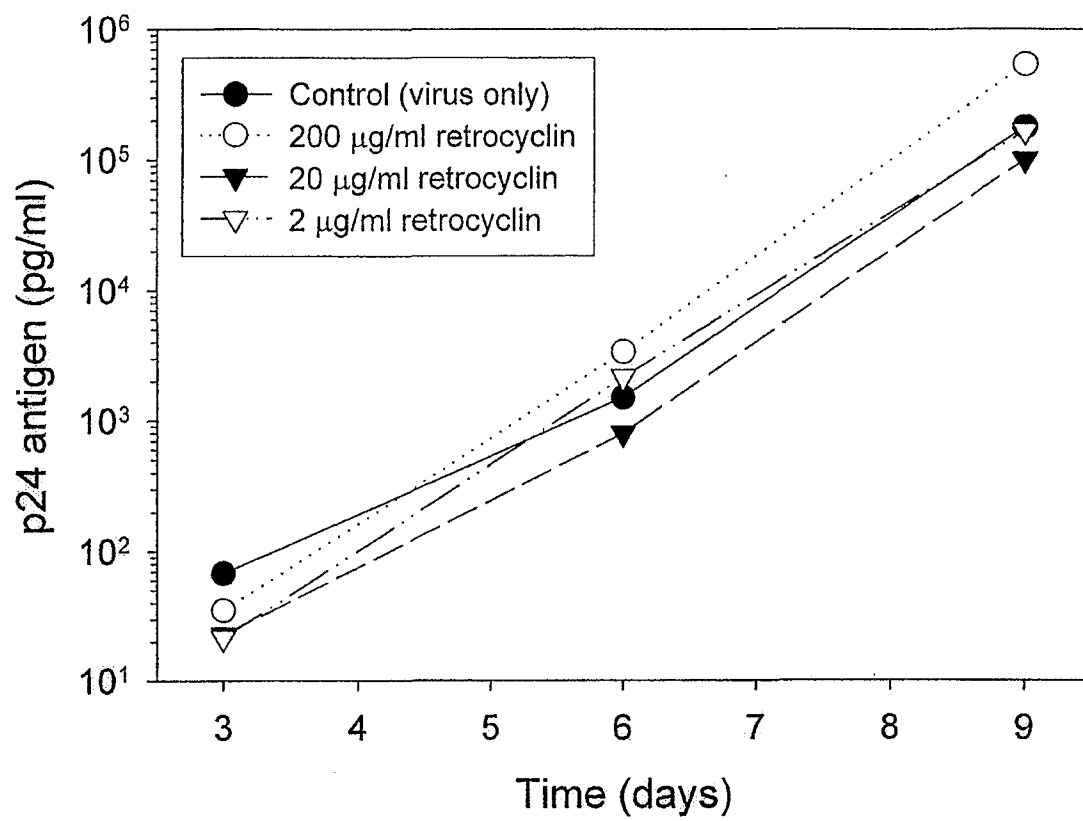


FIGURE 13

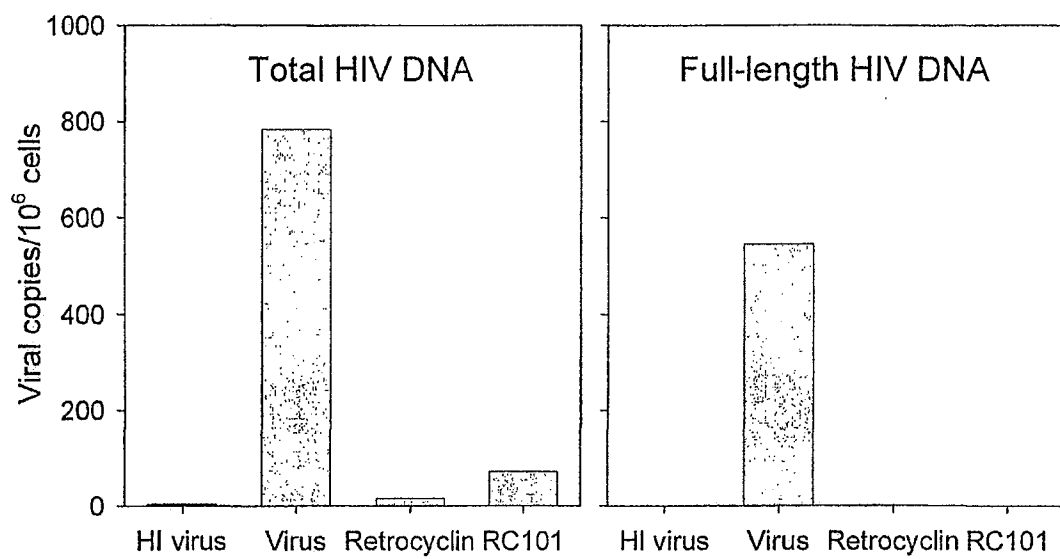


FIGURE 14

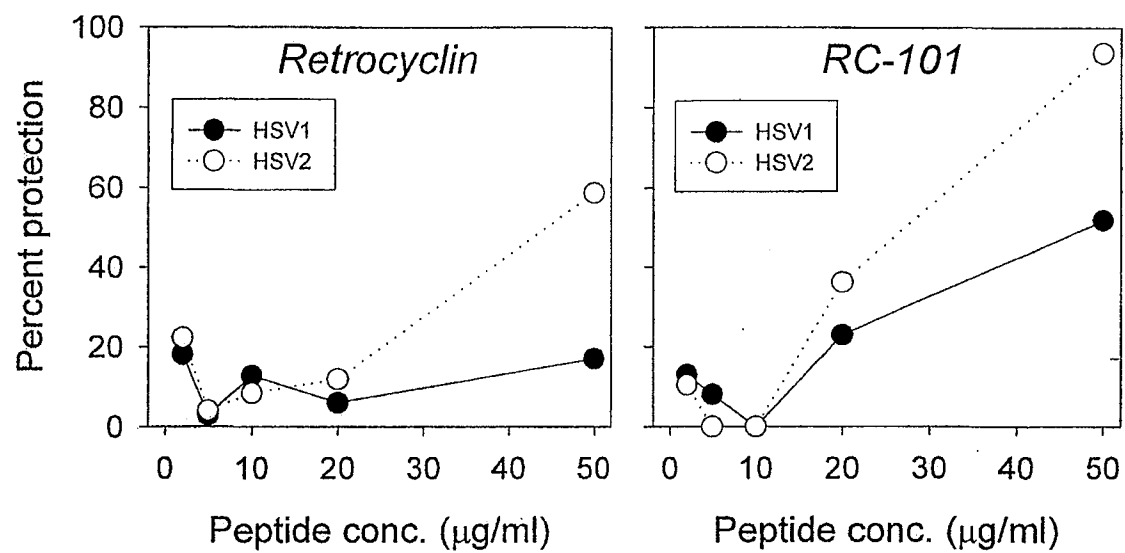


FIGURE 15 (Page 1)

Translation and Alignment of Human and Orangutan retrocyclin gene sequences

(SEQ ID NO 65) Human: VTPAMRTFALLTAMLLLVALL•AQAEPLQARADEAAAAQEQPGADDQEMAHAFTHWESAALPLSDSARGLRCTCGRGICRLL•RRFGSCA
 (SEQ ID NO 66) Orang 19: VTPAMRTFALLTAMLLLVALL•AEAEPLQARADETAAQEQPGADDQEMAHAFTHWDESATLPLSDSARGLRCTCRRGVCRRFL•RHLGSCA
 (SEQ ID NO 67) Orang 20: VTPAMRTFTVLAAAMLLLVVALQAQAEPLRARADETAAQEQPGADDQEMAHAFTHWDESAALPLSDSARGLRCTCRRGVCRRFL•RHLGSCA
 (SEQ ID NO 68) Orang 21: VTPAMRTFTVLAAAMLLLVVALQAQAEPLRARADETAAQEQPGADDQEMAHAFTHWDESAALPLSDSARGLRCTCRRGVCRRLL•RHFSGCA

Human: FRGTLHRICCR•ACRIKKHKLRIFYES•KKFLLLLLVVLHFLFSSKINTLLQDFSL (human)
 Orang 19: FRGTLHRICCR•ACRIKKKLRIFYES•KKFVFLLYLALHFLFSSKINTLLQDFCL (orangutan clone #19)
 Orang 20: FRGTLHRICCR•ACRIKKKLRIFYES•KKFVFLLYLALHFLFSSKINTLLQDFCL (orangutan clone #20)
 Orang 21: FRGTLHRICCR•ACRIKKKLRIFYES•KKFLLLYLALHFLFSSKINTLLQDFCL (orangutan clone #21)

15/16

Translation and Alignment of Human and Gorilla retrocyclin gene sequences

(SEQ ID NO 65) Human: VTPAMRTFALLTAMLLLVALL•AQAEPLQARADEAAAAQEQPGADDQEMAHAFTHWESAALPLSDSARGLRCTCGRGICRLL•RRFGSCA
 (SEQ ID NO 69) Gorilla VTPAMRTFALLTAMLLLVDL•AQAEPLQARADEAAAAQEQPGADDQEMAHAFTHWDESAALPLSDSARGLRCTCGRGICRLL•RRFGSCA

Human: FRGTLHRICCR•ACRIKKHKLRIFYES•KKFLLLLLVVLHFLFSSKINTLLQDFSL (human)
 Gorilla: FRGTLHRICCR•ACRIKKKLRIFYET•KKFLLLLLVVLHFLFSSKINTLLQDFCL (gorilla)

FIGURE 15 (Page 2)

Translation and Alignment of Human and Chimpanzee retrocyclin gene sequences

(SEQ ID NO 65) Human: VTPAMRTFALLTAMLLIVL•AQAEPLQARADEAAAQEQPGADDQEMAHAF•TWHEAALPLSDSARGLR•CICGRGICPLL•RRFGSCA
 (SEQ ID NO 70) P. troglodytes: VTPAMRTFALLTAMLLIVL•AQAEPLQARADEAAAQEQPGADDQEMAHAF•TWHEAALPLSDSARGLR•CICGRGICPLL•RRFGSCA
 (SEQ ID NO 71) P. paniscus: VTPAMRTFALLTAMLLIVL•AQAEPLQARADEAAAQEQPGADDQEMAHAF•TWHEAALPLSDSARGLR•CICGRGICPLL•RRFGSCA

Human: FRGTLHRICCR•ACRIKKHKLR•IYFES•KKFLLLLLYLVLHFLFSSKINTLLQDFSL (human)
 P. troglodytes: FRGTLHRICCR•ACRIKKHKLR•IYFES•KKFLLLLLYLVLHFLFSSKINTLLQDFSL (chimpanzee)
 P. paniscus: FRGTLHRICCR•ACRIKKHKLR•IYFES•KKFLLLLLYLVLHFLFSSKINTLLQDFSL (bonobo clone#6)

Alignment of Human, Rhesus monkey and pig-tailed Macaque sequences

(SEQ ID NO 65) VTPAMRTFALLTAMLLIVL•AQAEPLQARADEAAAQEQPGADDQEMAHAF•TWHEAALPLSDSARGLR•CICGRGICPLL•RRFGSCA
 (SEQ ID NO 72) VTPAMRTFALLTAMLLIVLHAQAEARQARADEAAAQEQPGADDQGMASHFTRPENNAALPLSESARGLR•CICRRGVCQLL•RRLGSCA
 (SEQ ID NO 73) VTPAMRTFALLTAMLLIVLHAQAEARQARADEAAAQEQPGADDQGMASHFTRPENNAALPLSESARGLR•CICRRGVCQLL•RRLGSCA

FRGTLHRICCR•ACRIKKHKLR•IYFES•KKFLLLLLYLVLHFLFSSKINTLLQDFSL (human)
 FRG•LCR•ICCR•ASRIKKNTLRSYFES•KKFLLLLLYLVLHFLFSSQINTFSQDFCL (rhesus monkey)
 FRG•LCR•ICCR•ASRIKKNTLRSYFES•KKFLLLLLYLVLHFLFSSQINTFSQDFCL (pig-tailed macaque clone #16)

SEQUENCE LISTING

<110> Robert Lehrer
Alan Waring
Alexander Cole
Teresa Hong

<120> Retrocyclins: Antiviral and
Antimicrobial Peptides

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<150> 60/284,855

<151> 2001-04-18

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Gly Arg

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Gly Arg

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 tctgtcacc cagccatgag gaccttcgcc ctctcactg ccatgcttct cctggtggcc 120
 ctg tag gct cag gcg gag cca ctt cag gca aga gct gat gaa gct gca 168
 * Ala Gln Ala Glu Pro Leu Gln Ala Arg Ala Asp Glu Ala Ala
 1 5 10
 gcc cag gag cag cct gga gca gat gat cag gaa atg gct cat gcc ttt 216
 Ala Gln Glu Gln Pro Gly Ala Asp Asp Gln Glu Met Ala His Ala Phe
 15 20 25 30

aca tgg cat gaa agt gcc gct ctt ccg ctt tca gac tca gcg aga ggc 264
 Thr Trp His Glu Ser Ala Ala Leu Pro Leu Ser Asp Ser Ala Arg Gly
 35 40 45

ttg agg tgc att tgc gga aga gga att tgc cgt ttg tta t aacgtcgctt 314
 Leu Arg Cys Ile Cys Gly Arg Gly Ile Cys Arg Leu Leu
 50 55

tgggtcctgc gcctttcgtg gtacactcca ccgcatctgc tgccgctgag cttgcagaat 374
 caagaaacat aagctcagaa ttacttttga gagttaaag aaattcttgt tactcctgta 434
 ccttgcctc catttccttt tctcatccaa aataaatacc ttgttgcaag atttctctct 494
 tt 496

<210> 12
 <211> 59
 <212> PRT
 <213> Homo sapiens

<400> 12
 Ala Gln Ala Glu Pro Leu Gln Ala Arg Ala Asp Glu Ala Ala Ala Gln
 1 5 10 15
 Glu Gln Pro Gly Ala Asp Asp Gln Glu Met Ala His Ala Phe Thr Trp
 20 25 30
 His Glu Ser Ala Ala Leu Pro Leu Ser Asp Ser Ala Arg Gly Leu Arg
 35 40 45
 Cys Ile Cys Gly Arg Gly Ile Cys Arg Leu Leu
 50 55

<210> 13
 <211> 97
 <212> PRT
 <213> Homo sapiens

<220>
 <221> PEPTIDE
 <222> (1)...(97)
 <223> Human defensin 4

<400> 13
 Met Arg Ile Ile Ala Leu Leu Ala Ala Ile Leu Leu Val Ala Leu Gln
 1 5 10 15
 Val Arg Ala Gly Pro Leu Gln Ala Arg Gly Asp Glu Ala Pro Gly Gln
 20 25 30
 Glu Gln Arg Gly Pro Glu Asp Gln Asp Ile Ser Ile Ser Phe Ala Trp
 35 40 45
 Asp Lys Ser Ser Ala Leu Gln Val Ser Gly Ser Thr Arg Gly Met Val
 50 55 60
 Cys Ser Cys Arg Leu Val Phe Cys Arg Arg Thr Glu Leu Arg Val Gly
 65 70 75 80
 Asn Cys Leu Ile Gly Gly Val Ser Phe Thr Tyr Cys Cys Thr Arg Val
 85 90 95
 Asp

<210> 14
 <211> 500
 <212> DNA
 <213> Macaca mulatta

<220>
 <221> CDS

<222> (95)...(325)
 <223> theta defensin 1A precursor

 <221> sig_peptide
 <222> (95)...(154)

 <221> mat_peptide
 <222> (287)...(313)
 <223> ligated to RTD1b in head-to-tail orientation to
 form the cyclic octadecapeptide RTD1; RTD1 is
 stabilized by three intramolecular disulfides

 <400> 14
 gacggctgct gttgctacag gagacccagg acagaggact gctgtctgca ctctctcttc 60
 actctgccta acttgaggat ctgtcactcc agcc atg agg acc ttc gcc ctc ctc 115
 Met Arg Thr Phe Ala Leu Leu
 -20 -15

 acc gcc atg ctt ctc ctg gtg gcc ctg cac gct cag gca gag gca cgt 163
 Thr Ala Met Leu Leu Val Ala Leu His Ala Gln Ala Glu Ala Arg
 -10 -5 1

 cag gca aga gct gat gaa gct gcc gcc cag cag cag cct gga aca gat 211
 Gln Ala Arg Ala Asp Glu Ala Ala Gln Gln Gln Pro Gly Thr Asp
 5 10 15

 gat cag gga atg gct cat tcc ttt aca tgg cct gaa aac gcc gct ctt 259
 Asp Gln Gly Met Ala His Ser Phe Thr Trp Pro Glu Asn Ala Ala Leu
 20 25 30 35

 cca ctt tca gag tca gcg aaa ggc ttg agg tgc att tgc aca cga gga 307
 Pro Leu Ser Glu Ser Ala Lys Gly Leu Arg Cys Ile Cys Thr Arg Gly
 40 45 50

 ttc tgc cgt ttg tta taa tgtcaccttg ggtcctgcgc ttttcgtggt 355
 Phe Cys Arg Leu Leu *
 55

 tgactccacc ggatctgctg ccgctgagct tccagaatca agaaaaatat gctcagaagt 415
 tactttgaga gttaaaagaa attcttgcta ctgctgtacc ttctcctcag tttccttttc 475
 tcatcccaaa taaatacctt atcgc 500

 <210> 15
 <211> 76
 <212> PRT
 <213> Macaca mulatta

 <220>
 <221> SIGNAL
 <222> (1)...(20)

 <400> 15
 Met Arg Thr Phe Ala Leu Leu Thr Ala Met Leu Leu Leu Val Ala Leu
 -20 -15 -10 -5
 His Ala Gln Ala Glu Ala Arg Gln Ala Arg Ala Asp Glu Ala Ala Ala
 1 5 10
 Gln Gln Gln Pro Gly Thr Asp Asp Gln Gly Met Ala His Ser Phe Thr
 15 20 25
 Trp Pro Glu Asn Ala Ala Leu Pro Leu Ser Glu Ser Ala Lys Gly Leu
 30 35 40
 Arg Cys Ile Cys Thr Arg Gly Phe Cys Arg Leu Leu
 45 50 55

<210> 16
 <211> 495
 <212> DNA
 <213> Macaca mulatta

 <220>
 <221> CDS
 <222> (90)...(320)
 <223> theta defensin 1b precursor

 <221> sig_peptide
 <222> (90)...(149)

 <221> mat_peptide
 <222> (282)...(308)
 <223> ligated to RTD1a in head-to-tail orientation to
 form the cyclic octadecapeptide RTD1; RTD1 is
 stabilized by three intramolecular disulfides

 <400> 16
 gaccgctgct cttgctacag gagacccggg acagaggact gctgtgtgcc ctctctcttc 60
 actctgccta acttgaggat ctgccagcc atg agg acc ttc gcc ctc ctc acc 113
 Met Arg Thr Phe Ala Leu Leu Thr
 -20 -15

 gcc atg ctt ctc ctg gtg gcc ctg cac gct cag gca gag gca cgt cag 161
 Ala Met Leu Leu Val Ala Leu His Ala Gln Ala Glu Ala Arg Gln
 -10 -5 1

 gca aga gct gat gaa gct gcc gcc cag cag cag cct gga gca gat gat 209
 Ala Arg Ala Asp Glu Ala Ala Ala Gln Gln Gln Pro Gly Ala Asp Asp
 5 10 15 20

 cag gga atg gct cat tcc ttt aca cgg cct gaa aac gcc gct ctt ccg 257
 Gln Gly Met Ala His Ser Phe Thr Arg Pro Glu Asn Ala Ala Leu Pro
 25 30 35

 ctt tca gag tca gcg aga ggc ttg agg tgc ctt tgc aga cga gga gtt 305
 Leu Ser Glu Ser Ala Arg Gly Leu Arg Cys Leu Cys Arg Arg Gly Val
 40 45 50

 tgc caa ctg tta taa aggcgtttgg ggctcctgcgc ttttcgtggt tgactctgcc 360
 Cys Gln Leu Leu *
 55

 ggatctgctg ccgctgagct tccagaatca agaaaaatac gctcagaagt tacttttgaga 420
 gttgaaagaa attcctgtta ctctgtacc ttgtcctcaa tttccttttc tcatcccaaa 480
 taaatacctt ctgcg 495

 <210> 17
 <211> 76
 <212> PRT
 <213> Macaca mulatta

 <220>
 <221> SIGNAL
 <222> (1)...(20)

 <400> 17
 Met Arg Thr Phe Ala Leu Leu Thr Ala Met Leu Leu Leu Val Ala Leu
 -20 -15 -10 -5
 His Ala Gln Ala Glu Ala Arg Gln Ala Arg Ala Asp Glu Ala Ala Ala
 1 5 10
 Gln Gln Gln Pro Gly Ala Asp Asp Gln Gly Met Ala His Ser Phe Thr

```

      15              20              25
Arg Pro Glu Asn Ala Ala Leu Pro Leu Ser Glu Ser Ala Arg Gly Leu
      30              35              40
Arg Cys Leu Cys Arg Arg Gly Val Cys Gln Leu Leu
      45              50              55

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<210> 18
 <211> 54
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> synthetic sequence

<400> 18
 aggtgcattt gcggaagagg aatttgcagg tgcatttgcg gaagaggaat ttgc 54

<210> 19
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> generated by replacement of variants in consensus
 sequence

<400> 19
 Arg Cys Ile Cys Gly Arg Gly Ile Cys
 1 5

<210> 20
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> generated by replacement of variants in consensus
 sequence

<400> 20
 Arg Cys Leu Cys Gly Arg Gly Ile Cys
 1 5

<210> 21
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> generated by replacement of variants in consensus
 sequence

<400> 21
 Arg Cys Ile Cys Arg Arg Gly Ile Cys
 1 5

<210> 22
 <211> 9
 <212> PRT
 <213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 22
Arg Cys Ile Cys Thr Arg Gly Ile Cys
1 5

<210> 23
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 23
Arg Cys Ile Cys Val Arg Gly Ile Cys
1 5

<210> 24
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 24
Arg Cys Ile Cys Gly Leu Gly Ile Cys
1 5

<210> 25
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 25
Arg Cys Ile Cys Gly Arg Gly Val Cys
1 5

<210> 26
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 26
Arg Cys Ile Cys Gly Arg Gly Phe Cys
1 5

<210> 27
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 27
Arg Cys Leu Cys Arg Arg Gly Val Cys
1 5

<210> 28
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 28
Arg Cys Leu Cys Thr Arg Gly Ile Cys
1 5

<210> 29
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 29
Arg Cys Leu Cys Val Arg Gly Ile Cys
1 5

<210> 30
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 30
Arg Cys Leu Cys Gly Leu Gly Val Cys
1 5

<210> 31
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
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sequence

<400> 31

Arg Cys Leu Cys Gly Arg Gly Val Cys
1 5

<210> 32

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 32

Arg Cys Leu Cys Gly Arg Gly Phe Cys
1 5

<210> 33

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 33

Arg Cys Ile Cys Arg Arg Gly Val Cys
1 5

<210> 34

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 34

Arg Cys Ile Cys Arg Arg Gly Phe Cys
1 5

<210> 35

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 35

Arg Cys Ile Cys Thr Arg Gly Val Cys
1 5

<210> 36

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 36

Arg Cys Ile Cys Thr Arg Gly Phe Cys
1 5

<210> 37

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 37

Arg Cys Ile Cys Thr Leu Gly Ile Cys
1 5

<210> 38

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 38

Arg Cys Ile Cys Val Leu Gly Phe Cys
1 5

<210> 39

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 39

Arg Cys Ile Cys Arg Leu Gly Ile Cys
1 5

<210> 40

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 40

Arg Cys Ile Cys Val Arg Gly Val Cys

1 5

<210> 41
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 41
Arg Cys Ile Cys Gly Arg Gly Phe Cys
1 5

<210> 42
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 42
Arg Cys Ile Cys Gly Leu Gly Phe Cys
1 5

<210> 43
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 43
Arg Cys Ile Cys Gly Leu Gly Val Cys
1 5

<210> 44
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 44
Arg Cys Leu Cys Arg Leu Gly Ile Cys
1 5

<210> 45
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 45
Arg Cys Leu Cys Arg Arg Gly Val Cys
1 5

<210> 46
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 46
Arg Cys Leu Cys Arg Arg Gly Phe Cys
1 5

<210> 47
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 47
Arg Cys Leu Cys Thr Leu Gly Ile Cys
1 5

<210> 48
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 48
Arg Cys Leu Cys Thr Arg Gly Val Cys
1 5

<210> 49
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 49
Arg Cys Leu Cys Thr Arg Gly Phe Cys
1 5

<210> 50
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 50
Arg Cys Leu Cys Val Leu Gly Ile Cys
1 5

<210> 51
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 51
Arg Cys Leu Cys Val Arg Gly Val Cys
1 5

<210> 52
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 52
Arg Cys Ile Cys Gly Arg Gly Ile Cys
1 5

<210> 53
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 53
Arg Cys Ile Cys Arg Leu Gly Val Cys
1 5

<210> 54
<211> 9
<212> PRT
<213> Artificial Sequence

<220>
<223> generated by replacement of variants in consensus
sequence

<400> 54

Arg Cys Ile Cys Arg Leu Gly Phe Cys
1 5

<210> 55

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 55

Arg Cys Ile Cys Thr Leu Gly Val Cys
1 5

<210> 56

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 56

Arg Cys Ile Cys Thr Leu Gly Phe Cys
1 5

<210> 57

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 57

Arg Cys Ile Cys Val Leu Gly Val Cys
1 5

<210> 58

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 58

Arg Cys Ile Cys Val Leu Gly Phe Cys
1 5

<210> 59

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 59

Arg Cys Leu Cys Gly Leu Gly Val Cys
1 5

<210> 60

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 60

Arg Cys Leu Cys Gly Leu Gly Ile Cys
1 5

<210> 61

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 61

Arg Cys Leu Cys Thr Leu Gly Val Cys
1 5

<210> 62

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 62

Arg Cys Leu Cys Thr Leu Gly Ile Cys
1 5

<210> 63

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus
sequence

<400> 63

Arg Cys Leu Cys Val Leu Gly Val Cys

1

5

<210> 64

<211> 9

<212> PRT

<213> Artificial Sequence

<220>

<223> generated by replacement of variants in consensus sequence

<400> 64

Arg Cys Leu Cys Val Leu Gly Ile Cys

1

5

<210> 65

<211> 140

<212> PRT

<213> Homo sapiens

<400> 65

Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Ala	Leu	Leu	Thr	Ala	Met	Leu	Leu
1				5					10					15	
Leu	Val	Ala	Leu	Ala	Gln	Ala	Glu	Pro	Leu	Gln	Ala	Arg	Ala	Asp	Glu
		20						25					30		
Ala	Ala	Ala	Gln	Glu	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Glu	Met	Ala	His
		35					40					45			
Ala	Phe	Thr	Trp	His	Glu	Ser	Ala	Ala	Leu	Pro	Leu	Ser	Asp	Ser	Ala
	50					55					60				
Arg	Gly	Leu	Arg	Cys	Ile	Cys	Gly	Arg	Gly	Ile	Cys	Arg	Leu	Leu	Arg
65					70					75				80	
Arg	Phe	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Thr	Leu	His	Arg	Ile	Cys	Cys
			85					90					95		
Arg	Ala	Cys	Arg	Ile	Lys	Lys	His	Lys	Leu	Arg	Ile	Tyr	Phe	Glu	Ser
		100						105					110		
Lys	Lys	Phe	Leu	Leu	Leu	Leu	Tyr	Leu	Val	Leu	His	Phe	Leu	Phe	Ser
		115					120					125			
Ser	Lys	Ile	Asn	Thr	Leu	Leu	Gln	Asp	Phe	Ser	Leu				
	130					135					140				

<210> 66

<211> 140

<212> PRT

<213> Orangutan

<400> 66

Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Ala	Leu	Leu	Ala	Ala	Met	Leu	Leu
1				5					10					15	
Leu	Val	Ala	Leu	Ala	Glu	Ala	Glu	Pro	Leu	Gln	Ala	Arg	Ala	Asp	Glu
		20						25					30		
Thr	Ala	Ala	Gln	Glu	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Glu	Met	Ala	His
		35					40					45			
Ala	Phe	Thr	Trp	Asp	Glu	Ser	Ala	Thr	Leu	Pro	Leu	Ser	Asp	Ser	Ala
	50					55					60				
Arg	Gly	Leu	Arg	Cys	Ile	Cys	Arg	Arg	Gly	Val	Cys	Arg	Phe	Leu	Arg
65					70					75				80	
His	Leu	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Thr	Leu	His	Arg	Ile	Cys	Cys
			85					90					95		
Arg	Ala	Cys	Arg	Ile	Lys	Lys	Asn	Lys	Leu	Arg	Ile	Tyr	Phe	Glu	Ser
		100						105					110		
Lys	Lys	Phe	Val	Phe	Leu	Leu	Tyr	Leu	Ala	Leu	His	Phe	Leu	Phe	Ser

Ser Lys Ile Asn Thr Leu Leu Gln Asp Phe Cys Leu
115 120 125
130 135 140

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<210> 67
<211> 141
<212> PRT
<213> Orangutan
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<400> 67															
Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Thr	Val	Leu	Ala	Ala	Met	Leu	Leu
1				5					10					15	
Val	Val	Ala	Leu	Gln	Ala	Gln	Ala	Glu	Pro	Leu	Arg	Ala	Arg	Ala	Asp
			20					25					30		
Glu	Thr	Ala	Ala	Gln	Glu	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Glu	Met	Ala
			35				40					45			
His	Ala	Phe	Thr	Trp	Asp	Glu	Ser	Ala	Ala	Leu	Pro	Leu	Ser	Asp	Ser
	50					55					60				
Ala	Arg	Gly	Leu	Arg	Cys	Ile	Cys	Arg	Arg	Gly	Val	Cys	Arg	Phe	Leu
65					70					75					80
<Arg	His	Leu	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Thr	Leu	His	Arg	Ile	Cys
				85					90					95	
Cys	Arg	Ala	Cys	Arg	Ile	Lys	Lys	Asn	Lys	Leu	Arg	Ile	Tyr	Phe	Glu
			100					105					110		
Ser	Lys	Lys	Phe	Val	Phe	Leu	Leu	Tyr	Leu	Ala	Leu	His	Phe	Leu	Phe
		115					120					125			
Ser	Ser	Lys	Ile	Asn	Thr	Leu	Leu	Gln	Asp	Phe	Cys	Leu			
	130					135					140				

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<210> 68
<211> 141
<212> PRT
<213> Orangutan
```

<400> 68															
Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Thr	Val	Leu	Ala	Ala	Met	Leu	Leu
1				5					10					15	
Val	Val	Ala	Leu	Gln	Ala	Gln	Ala	Glu	Pro	Leu	Arg	Ala	Arg	Ala	Asp
		20						25					30		
Glu	Thr	Ala	Ala	Gln	Glu	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Glu	Met	Ala
		35					40					45			
His	Ala	Phe	Thr	Trp	Asp	Glu	Ser	Ala	Ala	Leu	Pro	Leu	Ser	Asp	Ser
	50				55						60				
Ala	Arg	Gly	Leu	Arg	Cys	Ile	Cys	Arg	Arg	Gly	Val	Cys	Arg	Leu	Leu
65					70					75				80	
Arg	His	Phe	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Thr	Leu	His	Arg	Ile	Cys
				85					90					95	
Cys	Arg	Ala	Cys	Arg	Ile	Lys	Lys	Asn	Lys	Leu	Arg	Ile	Tyr	Phe	Glu
			100					105					110		
Ser	Lys	Lys	Phe	Leu	Phe	Leu	Leu	Tyr	Leu	Ala	Leu	His	Phe	Leu	Phe
		115				120						125			
Ser	Ser	Lys	Ile	Asn	Thr	Leu	Leu	Gln	Asp	Phe	Cys	Leu			
	130					135					140				

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<210> 69
<211> 140
<212> PRT
<213> Gorilla
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<400> 69
Val Thr Pro Ala Met Arg Thr Phe Ala Leu Leu Thr Ala Met Leu Leu

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1           5           10           15
Leu Val Asp Leu Ala Gln Ala Glu Pro Leu Gln Ala Arg Ala Asp Glu
20           25           30
Ala Ala Ala Gln Glu Gln Pro Gly Ala Asp Asp Gln Glu Met Ala His
35           40           45
Ala Phe Thr Trp Asp Glu Ser Ala Ala Leu Pro Leu Ser Asp Ser Ala
50           55           60
Arg Gly Leu Arg Cys Ile Cys Gly Arg Gly Ile Cys Arg Leu Leu Arg
65           70           75           80
Arg Phe Gly Ser Cys Ala Phe Arg Gly Thr Leu His Arg Ile Cys Cys
85           90           95
Arg Ala Cys Arg Ile Lys Lys Asn Lys Leu Arg Ile Tyr Phe Glu Thr
100          105          110
Lys Lys Phe Leu Leu Leu Tyr Leu Val Leu His Phe Leu Phe Ser
115          120          125
Ser Lys Ile Asn Thr Leu Leu Gln Asp Phe Cys Leu
130          135          140

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<210> 70
 <211> 141
 <212> PRT
 <213> Champanzee

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<400> 70
Val Thr Pro Ala Met Arg Thr Phe Ala Leu Leu Thr Ala Met Leu Leu
1           5           10           15
Leu Val Ala Leu Ala Gln Ala Glu Pro Leu Gln Ala Arg Ala Asp Glu
20           25           30
Ala Ala Ala Gln Glu Gln Pro Gly Ala Asp Asp Gln Glu Met Ala His
35           40           45
Ala Phe Thr Trp Asp Glu Ser Ala Ala Leu Pro Leu Ser Asp Ser Ala
50           55           60
Arg Gly Leu Arg Cys Ile Gly Gly Arg Gly Ile Cys Gly Leu Leu Gln
65           70           75           80
Arg Arg Phe Gly Ser Cys Ala Phe Arg Gly Thr Leu His Arg Ile Cys
85           90           95
Cys Arg Ala Cys Arg Ile Lys Lys Asn Lys Leu Arg Ile Tyr Ser Glu
100          105          110
Ser Lys Lys Phe Leu Leu Leu Leu Tyr Leu Val Leu His Phe Leu Phe
115          120          125
Ser Ser Lys Ile Asn Thr Leu Leu Gln Asp Phe Ser Leu
130          135          140

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 <213> Chimpanzee

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Leu Val Ala Leu Ala Gln Ala Glu Pro Leu Gln Ala Arg Ala Asp Glu
20           25           30
Ala Ala Ala Gln Glu Gln Pro Gly Ala Asp Asp Gln Glu Met Ala His
35           40           45
Ala Phe Thr Trp Asp Glu Ser Ala Ala Leu Pro Leu Ser Asp Ser Ala
50           55           60
Arg Gly Leu Arg Cys Ile Gly Gly Arg Gly Ile Cys Gly Leu Leu Gln
65           70           75           80
Arg Arg Val Gly Ser Cys Ala Phe Arg Gly Thr Leu His Arg Ile Cys
85           90           95
Cys Arg Ala Cys Arg Ile Lys Lys Asn Lys Leu Arg Ile Tyr Ser Glu

```

	100		105		110
Ser	Lys	Lys	Phe	Leu	Leu
	115		120		125
Ser	Ser	Lys	Ile	Asn	Thr
	130		135		140

<210> 72
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 <212> PRT
 <213> Rhesus monkey

<220>
 <221> VARIANT
 <222> 113
 <223> Xaa = Any Amino Acid

<400> 72

Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Ala	Leu	Leu	Thr	Ala	Met	Leu	Leu
1				5					10					15	
Leu	Val	Ala	Leu	His	Ala	Gln	Ala	Glu	Ala	Arg	Gln	Ala	Arg	Ala	Asp
		20						25				30			
Glu	Ala	Ala	Ala	Gln	Gln	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Gly	Met	Ala
		35					40					45			
His	Ser	Phe	Thr	Arg	Pro	Glu	Asn	Ala	Ala	Leu	Pro	Leu	Ser	Glu	Ser
	50					55					60				
Ala	Arg	Gly	Leu	Arg	Cys	Leu	Cys	Arg	Arg	Gly	Val	Cys	Gln	Leu	Leu
65					70					75				80	
Arg	Arg	Leu	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Leu	Cys	Arg	Ile	Cys	Cys
			85						90				95		
Arg	Ala	Ser	Arg	Ile	Lys	Lys	Asn	Thr	Leu	Arg	Ser	Tyr	Phe	Glu	Ser
			100						105				110		
Xaa	Lys	Lys	Phe	Leu	Leu	Leu	Leu	Tyr	Leu	Val	Leu	Asn	Phe	Leu	Phe
		115					120					125			
Ser	Ser	Gln	Ile	Asn	Thr	Phe	Ser	Gln	Asp	Phe	Cys	Leu			
		130					135				140				

<210> 73
 <211> 141
 <212> PRT
 <213> Pig-tailed macaque

<220>
 <221> VARIANT
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 <223> Xaa = Any Amino Acid

<400> 73

Val	Thr	Pro	Ala	Met	Arg	Thr	Phe	Ala	Leu	Leu	Thr	Ala	Met	Leu	Leu
1				5					10					15	
Leu	Val	Ala	Leu	His	Ala	Gln	Ala	Glu	Ala	Arg	Gln	Ala	Arg	Ala	Asp
		20						25				30			
Glu	Ala	Ala	Ala	Gln	Gln	Gln	Pro	Gly	Ala	Asp	Asp	Gln	Gly	Met	Ala
		35					40					45			
His	Ser	Phe	Thr	Arg	Pro	Glu	Asn	Ala	Ala	Leu	Pro	Leu	Ser	Glu	Ser
	50					55					60				
Ala	Arg	Gly	Leu	Arg	Cys	Ile	Cys	Arg	Arg	Gly	Val	Cys	Gln	Leu	Leu
65					70					75				80	
Arg	Arg	Leu	Gly	Ser	Cys	Ala	Phe	Arg	Gly	Leu	Cys	Arg	Ile	Cys	Cys
			85						90				95		
Arg	Ala	Ser	Arg	Ile	Lys	Lys	Asn	Thr	Leu	Arg	Ser	Tyr	Phe	Glu	Ser
			100						105				110		
Xaa	Lys	Lys	Phe	Leu	Leu	Leu	Leu	Tyr	Leu	Val	Leu	Asn	Phe	Leu	Phe

		115					120				125	
Ser	Ser	Gln	Ile	Asn	Thr	Phe	Ser	Gln	Asp	Phe	Cys	Leu
	130						135				140	

INTERNATIONAL SEARCH REPORT

PCT/US02/12353

Box I Observations where certain claims were found unsearchable 1. because they relate to subject matter not required to be searched by this Authority, namely:

Claim 7 is unsearchable because no meaningful search can be run on a peptide sequence wherein there are no static peptide residues. While the applicant seems to be trying to claim a consensus sequence, the applicant has not identified any one sequence that the examiner can search within claim 7. Although claims 8-10 depend from the unsearchable claim 7, the examiner feels that they resolve the problem sufficiently that they may be meaningfully examined even though the generic claim is not searchable. Claim 16 is unsearchable as being an improperly dependent claim. It is a claim further limiting a method. However, the claim from which it depends is a product claim, not a method. Therefore, no meaningful search may be made of claim 16.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING

Group I. Claims 1-6, 8-11, and 15, drawn to isolated retrocyclin peptides and a method of using the peptides to prevent retroviral infection of a cell by administering the peptide to the cell.

Group II. Claims 12-14, drawn to isolated nucleic acids encoding retrocyclin.

Group III. Claim 17, drawn to a method of killing microbial organisms by administering the peptide to the microbial organism.

Group IV. Claim 18, drawn to a method of administering a retrocyclin as a therapeutic agent to a patient with a microbial infection.

Group V. Claim 18, drawn to a method of administering a retrocyclin as a therapeutic agent to a patient with a viral infection.

Group VI. Claim 19, drawn to a method of administering a retrocyclin as a prophylactic to prevent microbial infection of a patient at risk of such an infection.

Group VII. Claim 19, drawn to a method of administering a retrocyclin as a prophylactic to prevent viral infection of a patient at risk of such an infection.

Groups I and II comprise multiple inventions as defined below. If inventions from either of these Groups are elected for searching, the applicant must also elect which inventions within these Groups the applicant wishes to have examined. The inventions within these are set forth below.

Group I comprises multiple inventions, each of which comprises a different retrocyclin peptide. Each of the peptides is identified either by a combination of the sequences of sequence identification numbers 19-64, or by one of the sequences of 1-10. Thus, if the applicant wishes other sequences other than a peptide comprising two linked nonapeptides of SEQ ID NO: 19 (the first named inventions), the applicant must elect both Group I and either two sequences of SEQ ID NO: 19-64, or one of sequences 1-10.

Group II comprises multiple inventions, each of which is a nucleic acid encoding for a different peptide. If the applicant wished to elect a nucleotide encoding a retrocyclin for searching, the applicant must elect Group V, and one of the peptide sequences of SEQ ID NO: 1-10.

The inventions within Group I do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: each of the inventions within Group I has a different structure from the other polypeptides in the Group. They each have a unique structure (sequence) from the other polypeptides. As each of the polypeptides has a different structure, they share no special technical features one with another.

The inventions of Group II do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: each of the nucleic acids encodes for a different polypeptide. As each of the nucleic acids encodes a different polypeptide, they share no special technical feature.

The inventions listed as Groups I, and III-VII and as Group II above do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: the two Groups of invention relate to molecules with different structures and which perform different functions. Groups I and III-VII relate to polypeptides while Group II relates to nucleic acid sequences. Each of the two molecules is comprised of a different type of constituents, amino acids and nucleic acids respectively. Further, each molecule performs a different function- the peptides are useful in preventing or inhibiting infections, and in killing microbes, while the nucleic acid is used to encode for and produce proteins or peptides. As the two types of molecules have different structures and different functions, they share no common special technical feature. Therefore, they are not a part of the same general inventive concept.

The inventions listed as Groups I, and III-VII do not relate to a single general inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: each of the Groups comprises a different method of use for the claimed polypeptides. Under PCT Rule 13.1, the applicant is entitled to joinder of a

INTERNATIONAL SEARCH REPORT

PCT/US02/12353

product and a first method of using that product. The applicant has included further methods of using the product that produce different effects. Because of these different effects, these methods do not share a common special technical feature with the first method of use. As a first method of use is joined with the product, the remaining methods of use do not form a part of the same general inventive concept as the product and the first method of use.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US02/12353

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claim Nos.: 7 and 16
because they relate to subject matter not required to be searched by this Authority, namely:
Please See Continuation Sheet
2. ☐ Claim Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☐ Claim Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:
Please See Continuation Sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-6, 8, 10, 11, and 15

Remark on Protest

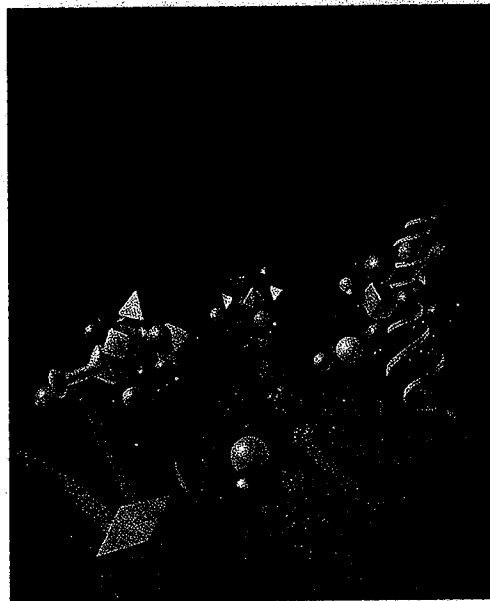
☐
☐

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

EXHIBIT 2

MOLECULAR CELL



BIOLOGY

T H I R D E D I T I O N

Harvey Lodish

David Baltimore

Arnold Berk

S. Lawrence Zipursky

Paul Matsudaira

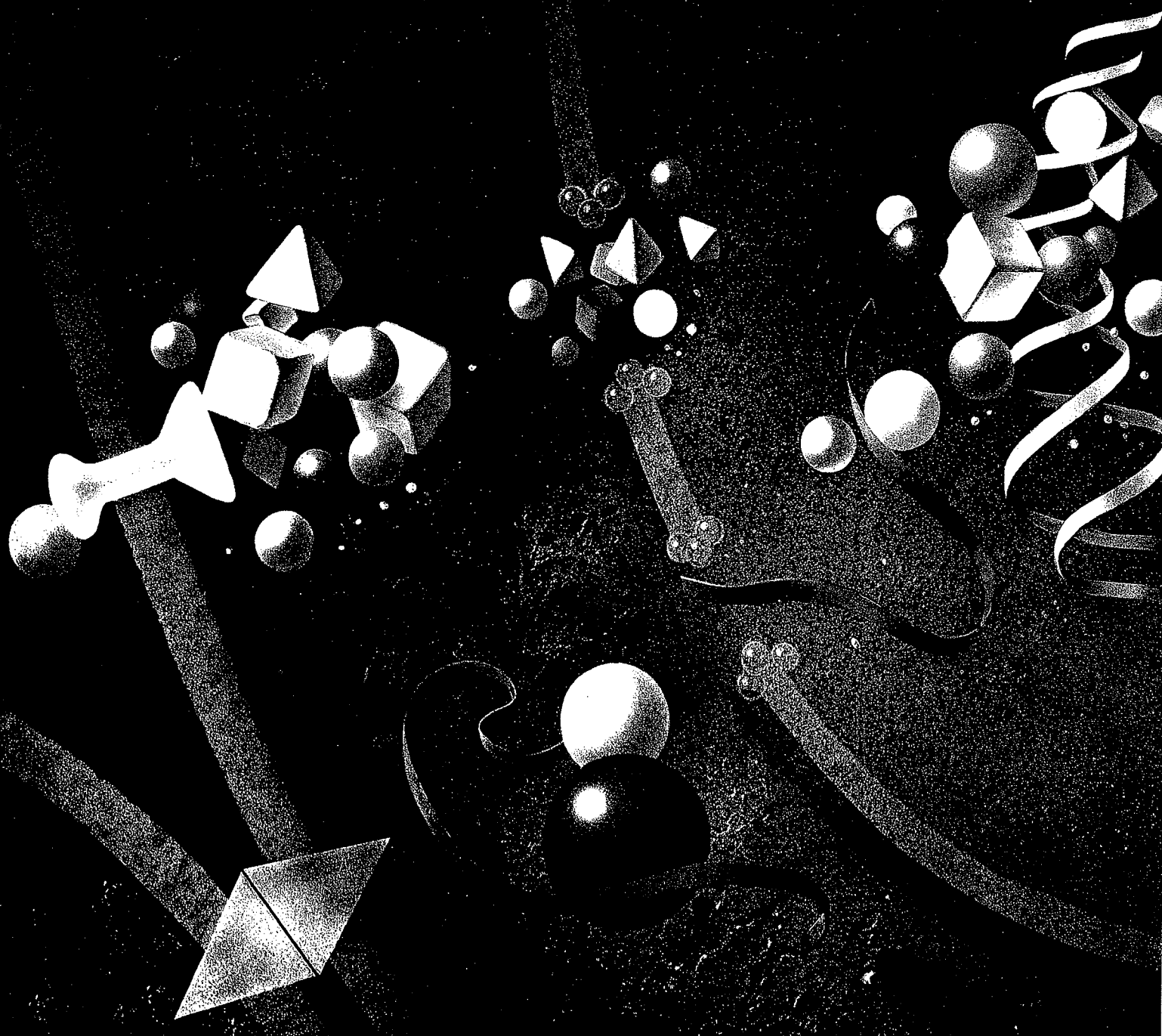
James Darnell

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T H I R D E D I T I O N

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ARNOLD BERK • S. LAWRENCE ZIPURSKY
PAUL MATSUDAIRA • JAMES DARNELL



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